

**BUKTI KORESPONDENSI**  
**SYARAT KHUSUS ARTIKEL JURNAL BEREPUTASI**

Judul Artikel : Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield

Jurnal : Journal of Ecological Engineering, Vol 24, No. 8, Tahun 2023, Hal. 128-136

Penulis : Zulkifli Maulana, Andi Muhibuddin, Muhammad Arief Nasution, Abri, Amiruddin Amirudin, Baharuddin, Andi Tenri Fitriyah, Sitti Nurani Sirajuddin, Abdel Razzaq M. Al Tawaha

No	Perihal	Tanggal
1	Registrasi akun ke Journal of Ecological Engineering	18 April 2023
2	Konfirmasi akun	18 April 2023
3	Submit artikel ke Journal of Ecological Engineering	10 Mei 2023
4	Konfirmasi Review dan Hasil Review	17 Mei 2023
5	Pengiriman revisi artikel ke Journal of Ecological Engineering	18 Mei 2023
6	Konfirmasi penerimaan revisi penulis ke editor	19 Mei 2023
7	Konfirmasi penerimaan artikel	29 Mei 2023
8	Konfirmasi proof version	19 Juni 2023

**1. REGISTRASI AKUN KE JOURNAL OF ECOLOGICAL ENGINEERING  
18 APRIL 2023**



Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

---

## Editorial System registration to Journal of Ecological Engineering

---

Journal of Ecological Engineering <office@jeeng.net>

18 April 2023 pukul 13.48

Kepada: muhibuddin <muhibuddin@universitasbosowa.ac.id>, Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

Dear Authors,

Thank you for your registration to the Editorial System of Journal of Ecological Engineering

Following confirmation, you will be able to log in to the Editorial System available here:

<https://www.editorialsystem.com/jeeng/>

Editorial Office of Journal of Ecological Engineering

**2. KONFIRMASI AKUN**

**18 APRIL 2023**



Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

---

## Editorial System account confirmation

---

Journal of Ecological Engineering <office@jeeng.net>

18 April 2023 pukul 15.23

Kepada: muhibuddin <muhibuddin@universitasbosowa.ac.id>, Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

Dear Authors,

After this confirmation you can log in to the system available here:

<https://www.editorialsystem.com/jeeng/>

Editorial Office of Journal of Ecological Engineering

**3. SUBMIT ARTIKEL KE JOURNAL OF ECOLOGICAL ENGINEERING  
10 MEI 2023**



Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

---

**New manuscript received by Editorial Office (JEENG-04241-2023-01)**

---

Journal of Ecological Engineering <office@jeeng.net>

10 Mei 2023 pukul 02.32

Kepada: muhibuddin <muhibuddin@universitasbosowa.ac.id>, Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

Dear Authors,

Thank you for your manuscript: Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield. The Following number has been assigned to it: JEENG-04241-2023-01.

The Manuscript will be checked by Editor and then sent to the Reviewers.  
You will be informed by email about any further decision on this article.

Thank you for submitting your work to our journal.

Kindest regards,  
Prof. Gabriel Borowski  
Editor-in-Chief  
Journal Of Ecological Engineering

**4. KONFIRMASI REVIEW DAN HASIL REVIEW**  
**17 MEI 2023**





Zulkifli Maulana &lt;zulkifli.maulana@universitasbosowa.ac.id&gt;

---

**Decision on manuscript JEENG-04241-2023-01**

---

Journal of Ecological Engineering &lt;office@jeeng.net&gt;

17 Mei 2023 pukul 13.27

Kepada: muhibuddin &lt;muhibuddin@universitasbosowa.ac.id&gt;, Zulkifli Maulana &lt;zulkifli.maulana@universitasbosowa.ac.id&gt;

May 17, 2023

JEENG-04241-2023-01

Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield

Dear Authors,

I am pleased to inform you that your manuscript, entitled: Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield, might be accepted for publication in our journal, pending some minor changes suggested by reviewers.

Please revise your paper strictly according to the attached Reviewers comments. Your manuscript won't be taken into consideration without the revisions made according to the recommendations.

Authors of our journal are requested to prepare a revised version of their manuscript as soon as possible. This may ensure fast publication if an article is finally accepted.

Thank you for submitting your work to us.

Kindest regards,  
Justyna Kujawska  
Managing Editor  
Journal of Ecological Engineering



## REVIEWERS COMMENTS

Journal of Ecological Engineering <office@jeeng.net>

17 Mei 2023 pukul 16.48

Kepada: muhibuddin <muhibuddin@universitasbosowa.ac.id>, Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

Please complete the form and return to the Editor

### A. MANUSCRIPT

Journal	Journal Of Ecological Engineering
Manuscript Number	JEENG-04241-2023-01
Title Of Paper	Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield

### B. REVIEWER'S SPECIFIC COMMENTS PER SECTION OF MANUSCRIPT

Title	The title is mostly appropriate
Abstract	The authors should correct all grammatical, spellings and typo
Keywords	Keywords are appropriate
Introduction	The authors should briefly discuss the relevance of the research methods to the research. Authors should correct all grammatical, spellings, italic and typo.
Materials and Methods	The authors should improve the table to make it easy to read. Authors should clarified using what statistical analysis. All grammatical, spellings and typo should be corrected.
Results and Discussion	The authors should improve the table to make it easy to read. All grammatical, spellings, italic and typo should be corrected.
Conclusion	Conclusion are appropriate
References	The authors should delete brackets in whole reference

## INTRODUCTION

Indonesia's soybean demand continues to increase along with population growth. This is due to the need for soybeans in the community for consumption, so the increase in soybean production is one indicator of the success of the agricultural sector. The high demand for soybeans is because it is one of the basic needs and the main source of vegetable protein for most Indonesians. In addition, soybeans are a group of leguminous plants that can fix nitrogen from the air that occurs in root nodules (Al-Tawaha et al., 2022a; Al-Tawaha et al., 2022b; Saranraj et al., 2021; Shiri Janagard and Ebadi Segherloo, 2015) with an average protein content of 40% based on dry matter (Al-Tawaha and Al-Tawaha, 2017; Cruz-Suárez et al., 2009; Banaszkiwicz, 2011), soybean consumption in Indonesia is 2.2 million tons per year. This figure is far from the average production level of only 819,442 tons per year. As a result, the government must import soybeans as much as 2–2.6 million tons per year. Domestic soybean production is currently only able to meet about 35% of the consumption (Ningrum et al., 2018). The governments have made several attempts to increase domestic soybean production, such as by clearing new farming land. Ultisol soil is one of the alternative lands for developing soybean areas in Indonesia, whose distribution reaches 45,794,000 ha or about 25% of the total land area of Indonesia (Syahputra et al., 2015), with great potential to be developed into productive land. However, ultisol soils are poor in organic matter (<5%), low pH, high Al saturation, low base saturation, and organic C, causing nutrient deficiencies, especially molybdenum (Mo) (Fitriati et al., 2014). On the other hand, the element Mo is needed in the

**Table 1.** Chemical and physical properties of ultisol soils before and after treatments

Parameters	Content before treatment (R <sub>0</sub> )	Content after treatment (R <sub>1</sub> )	Content after treatment (R <sub>2</sub> )	Content after treatment (R <sub>3</sub> )
pH (1:2,5) H <sub>2</sub> O	4,9	5,50	5,65	5,70
C-Organik (%)	0,82	0,83	0,83	0,84
N-Total (%)	0,18	0,28	0,36	0,38
Available-P (ppm)	10,00	10,20	10,37	10,48
K-dd (me/100 g)	0,32	0,43	0,47	0,51
CEC (me/100 g)	15,07	15,10	16,10	16,20
Base Saturation (%)	15,15	16,10	16,12	16,40
Al-dd (me/100 g)	1,92	10,20	10,20	10,20
Mo (ppm)	0,35	1,51	1,69	2,10
Al Saturation	22,64	22,12	21,78	21,60
Fraktion: Sand (%)	55,23	55,21	55,21	55,10
Dust (%)	24,10	24,12	24,13	24,32
Clay (%)	20,67	20,67	20,66	20,58
Tekstur	(Sandy clay loam)	(Sandy clay loam)	(Sandy clay loam)	Sandy clay loam)

## Observation and measurement

Vegetative growth observation included plantheight, age of flower appearance, number of root nodules, percentage of effective root nodules, and nitrogen (N) and potassium (K) uptake. Meanwhile, observations for the generative phase were the weight of 100 seeds, the number of pods per plant, and dry seed weight per plant.

## Data analysis

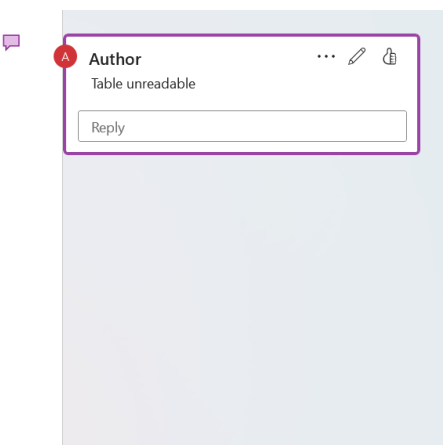
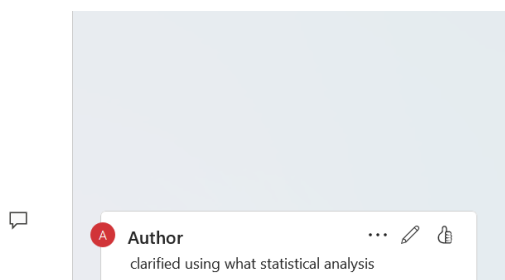
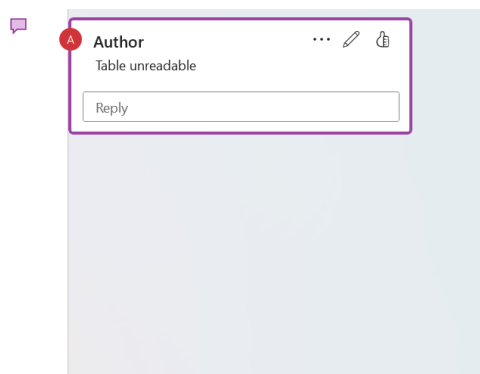
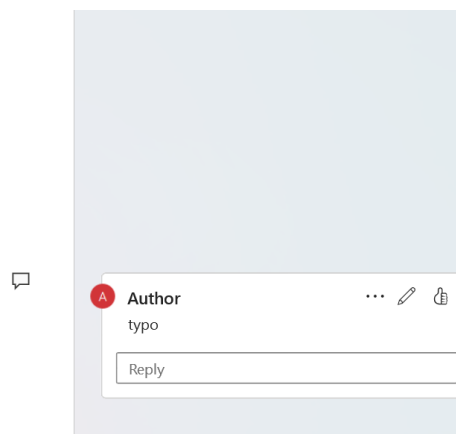
The data from the experiment were statistically analyzed. The differences among treatments were compared using analysis of variance (ANOVA) with the least significant difference (LSD) posthoc test at a 5% probability level.

**Table 2.** Average plant height, age of flower appearance, number of root nodules, and percentage of effective root nodules on soybean cultivars and the combinations of *Rhizobium* strain *Nod-Fix*<sup>+</sup>, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate

Treatment	Plant height (cm)	Age of flower appearance (days)	Number of root nodules (seeds)	Percentage of effective root nodules (%)
<b>Cultivars</b>				
V1	83.96 <sup>x</sup>	32.78 <sup>x</sup>	40.22 <sup>x</sup>	62.21 <sup>x</sup>
V2	75.62 <sup>y</sup>	38.96 <sup>y</sup>	34.64 <sup>y</sup>	58.28 <sup>y</sup>
<b>Sub-plots</b>				
R0	66.04 <sup>a</sup>	35.38 <sup>a</sup>	09.80 <sup>a</sup>	27.23 <sup>a</sup>
R1	82.59 <sup>b</sup>	36.05 <sup>a</sup>	26.14 <sup>b</sup>	67.76 <sup>b</sup>
R2	90.16 <sup>c</sup>	35.66 <sup>a</sup>	57.22 <sup>cd</sup>	79.76 <sup>cd</sup>
R3	80.37 <sup>b</sup>	36.10 <sup>a</sup>	48.47 <sup>c</sup>	73.74 <sup>c</sup>

Note: Numbers followed by the same letter in each column are not significantly different at Duncan's test level = 0.05

The results of Duncan's test in Table 2 on the observation of the number of root nodules showed that different treatments gave different results on the number of root nodules. Treatment R<sub>3</sub> showed the highest number of root nodules. This was due to the suitability of the needs for bacterial strain *Nod-Fix*<sup>+</sup>, Lime (CaCO<sub>3</sub>), Molybdenum (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O, and Cobalt (CoCl<sub>2</sub>). Therefore, metabolic processes



The observation results of the distribution pattern of the root nodules in Table 2 showed the best treatment of R<sub>2</sub>. The nodule distribution pattern determines the effectiveness of the nodule. Nodules scattered on the root neck indicate nodules effective in fixing N. The distribution pattern of root nodules explains the effectiveness of root nodules in fixing nitrogen free from the air. Effective root nodules are located on the taproots, while ineffective root nodules are scattered on lateral roots or root branches and are pale in color. The activity of N<sub>2</sub> fixation by bacteria is essential for overall N balance because the continuous form of fixed N can be lost due to denitrification and permeation (Howie & Echandi, 1983).

#### Nitrogen, phosphorus and potassium uptake

The ANOVA results showed that cultivars (V) had a significant effect on nitrogen (N) and potassium (K) uptake but had no significant effect on Phosphorus (P) uptake. The combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate (R) significantly affected N, P, and K up-take (Table 3). There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate. The *Baluran* cultivar (V1) produced higher N and K uptake, significantly different from the Willis cultivar (V2). However, the P uptake of treatments was not significantly different. According to

Table 3 shows that N, P, and K uptake increased with increasing doses of the combinations (CaCO<sub>3</sub> + NH<sub>4</sub>-molybdate). The highest increase was obtained in the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/ha of 852.7 mg/plant, 52.4 mg/plant, and 442.9 mg/plant, respectively. However, the results for the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/ha were decreased statistically significantly. The increased uptake of N, P, and K was thought to be the result of the ability of soybean cultivars to form root nodules in response to *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* inoculation. In addition, it was supposed that the number of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* in that treatment was more suitable than others. The root nodules were formed if there was a suitability between *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* and its host.

The results of Duncan's test in Table 3 showed that the highest nitrogen uptake was found in the R<sub>2</sub> treatment compared to other treatments (R<sub>0</sub>, R<sub>1</sub> and R<sub>3</sub>). The significant effect of the R treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* + MoCo (1.0: 0.6) kg/ha) was thought to result from the soybean's ability to form root nodules in response to the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* inoculation. In addition, it is suspected that the number of bacterial strains *Nod<sup>+</sup> Fix<sup>+</sup>* in this treatment is more suitable than the other treatments, and effective root nodules would form if there were a suitability between the bacteria and its host.

Table 3. Average nutrient uptake of N, P and K in soybean cultivars and combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate

Treatment	nutrient uptake (mg/plant)		
	N	P	K
<b>Cultivars</b>			
V1	710.6 <sup>a</sup>	43.1 <sup>a</sup>	370.0 <sup>a</sup>
V2	641.1 <sup>b</sup>	40.5 <sup>a</sup>	364.7 <sup>b</sup>
<b>Sub-plots</b>			
R0	385.6 <sup>a</sup>	33.1 <sup>a</sup>	244.8 <sup>a</sup>
R1	737.3 <sup>b</sup>	41.2 <sup>b</sup>	377.3 <sup>b</sup>
R2	852.7 <sup>c</sup>	52.4 <sup>c</sup>	442.9 <sup>bc</sup>
R3	727.8 <sup>b</sup>	40.6 <sup>b</sup>	405.4 <sup>b</sup>

Note: numbers followed by the same letter in each column are not significantly different at Duncan's test level = 0.05.

#### Leaf color scale

The observation results of the leaf color scale in Table 4 showed that the treatment of bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* with Mo and CaCO<sub>3</sub> lime supplements on soybeans using ultisol soils affected the leaf color scale. Table 4 shows that the Willis and *Baluran* varieties treated with the *Nod<sup>+</sup> Fix<sup>+</sup>* bacterial strain, Mo and CaCO<sub>3</sub> lime supplements had various leaf color scales.

Table 4. Leaf Color Scale.

Varietas	Treatments	leaf color scales. *)			
		1	2	3	4
V <sub>1</sub>	R <sub>0</sub>	x			
	R <sub>1</sub>		x		
	R <sub>2</sub>			x	
	R <sub>3</sub>			x	
V <sub>2</sub>	R <sub>0</sub>	x			
	R <sub>1</sub>				x
	R <sub>2</sub>			x	
	R <sub>3</sub>			x	

Note: \*) Criteria based on color scale, diagnosis of nutrient status from leaf color. The strengthening of legumes in relation to cropping system research project, Japan International Cooperation Agency (JICA): 1 – yellowish green, 2 – light green, 3 – green, 4 – slightly dark green.

Table 4 showed that the results on the leaf color scale was the best treatment of R<sub>1</sub> for the *Baluran* cultivar compared to other treatments. This showed that the fixed N element was used to meet the N required by plants so the availability of N for plants increases. An effective fixation, 50–75% of the total N requirement by plants can be fulfilled. In general, this study showed the pattern of increasing plant height, root dry weight, number of root nodules, number of red nodules, N uptake, root nodule distribution pattern, leaf color scale, weight of 100 seeds, and dry seed yield. It was found that the treatment of the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* with supplements of microelements (MoCo), lime (CaCO<sub>3</sub>), and Cobalt, simultaneously increased the availability of Mo in the soil so as to increase the uptake of Mo by plants. Jaiswal et al. (2021) stated that the element Mo functions as an activator of nitrogenase and reducing enzymes which activate *Rhizobium* performance in the N fixation process so that N binding increases. Nitrogen can stimulate cell elongation, vegetative growth, enlarge pods, slow down the initiation phase, and increase seeds yield and content (Ohyama et al., 2013). The results of Duncan's test in Table 4 showed that the highest weight of 100 seeds was obtained in the R treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* + MoCo (1.0: 0.6) kg/ha) compared to other treatments (R<sub>0</sub>, R<sub>1</sub> and R<sub>3</sub>). This revealed that the treatment is vital for the growth and production of soybeans.

Author  
typo

Reply

Author  
Italic and typo

Reply

Author  
Table unreadable

Reply

Author  
Table unreadable

Reply

Author  
typo

Reply

**Table 5.** The average weight of 100 seeds, number of pods per plant, and dry seed weight per plant

Treatment	Weight 100 seeds (g)	Number of pods per plant (piece)	Dry seed weight per plant (g)
<b>Cultivars</b>			
V1	13,75 <sup>a</sup>	28,66 <sup>a</sup>	16,53 <sup>a</sup>
V2	11,34 <sup>a</sup>	26,51 <sup>a</sup>	15,17 <sup>a</sup>
<b>Sub-plots</b>			
R0	10,37 <sup>a</sup>	19,18 <sup>a</sup>	10,99 <sup>a</sup>
R1	14,04 <sup>b</sup>	26,94 <sup>b</sup>	17,97 <sup>bc</sup>
R2	14,49 <sup>b</sup>	35,89 <sup>c</sup>	19,45 <sup>c</sup>
R3	11,88 <sup>a</sup>	28,34 <sup>b</sup>	15,00 <sup>b</sup>

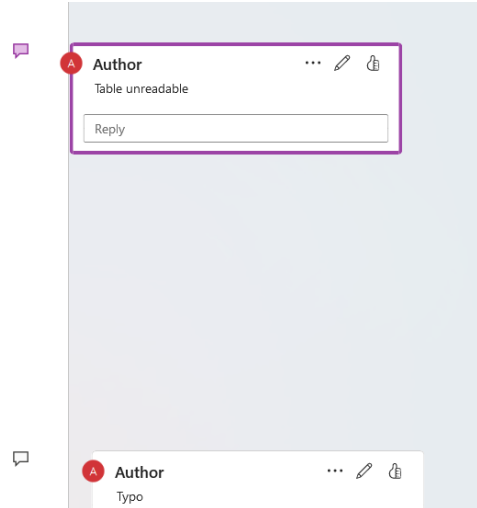
Note: numbers followed by the same letter in each column are not significantly different at Duncan's test level = 0.05.

This was due to the Cofactor Mo (MoCo) forming the active site of all eukaryotic molybdenum enzymes.

#### Yield components

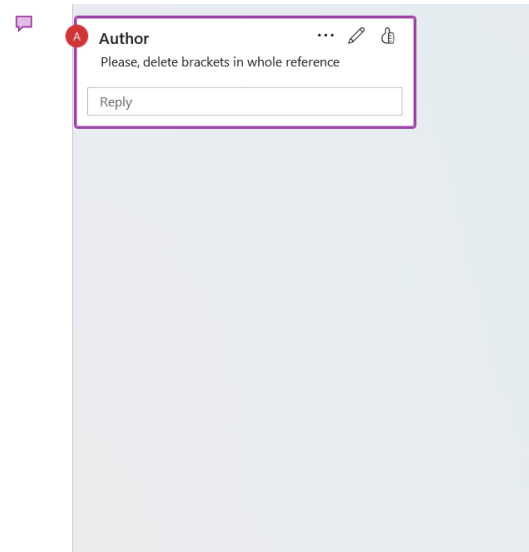
The ANOVA results showed that cultivars (V) had no significant effect; while the combinations of *Rhizobium* strain *Nod:Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub> – molybdate had a significant effect on the weight of 100 seeds, number of pods per plant, and dry seed weight per plant. There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod:Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub> – molybdate (Table 5).

Table 5 shows that the weight of 100 seeds, number of pods per plant, and dry seed weight per plant were



#### REFERENCES

1. Al-Tawaha, A.R.M., McNeil, D., Othman, Y.A., AlRawashdeh, I.M., Adnan, M., Zamin, M., Qaisi, A.M., Al-Tawaha, A., Jahan, N., Shah, M.A., Khalid, S. (2022a) Legume Production and Climate Change. In Improvement of Plant Production in the Era of Climate Change. CRC Press, 221–248.
2. Al-Tawaha, A.R.M.S., Khanum, S., Benkeblia, N., Khalid, S., Al-Tawaha, A.R., Mondal, M., Odat, N., Dey, A., Alimad, N., Thangadurai, D., Sangeetha, J. (2022b). Adapting Crops to Climate Change. Climate Change and Agriculture: Perspectives, Sustainability and Resilience, 53–77. <https://doi.org/10.1002/9781119789789.ch3>
3. Al-Tawaha, A.R., Al-Tawaha, A.R.M. (2017). Response of soybean plants to exogenous application of yeast extract: Growth and chemical composition. American-Eurasian Journal of Sustainable Agriculture, 11(2), 31–36.
4. Banaszkiwicz, T. (2011). Nutritional value of soybean meal. Soybean and Nutrition, 1–21. <https://doi.org/10.5772/23306>
5. Bhattacharya, P.T., Misra, S.R., Hussain, M. (2016). Nutritional aspects of essential trace elements in oral health and disease: an extensive review. Scientifica, 1–12. <https://doi.org/10.1155/2016/5464373>
6. Ciesła, J., Kopycińska, M., Łukowska, M., Bieganski, A., Janczarek, M. (2016). Surface properties of wild-type *Rhizobium leguminosarum* by Trifolij strain 24.2 and Its derivatives with different extracellular polysaccharide content. PLoS ONE, 11(10), 1–21. <https://doi.org/10.1371/journal.pone.0165080>
7. Cruz-Suárez, L.E., Tapia-Salazar, M., Villarreal-Cavazos, D., Beltran-Rocha, J., Nieto-López, M.G., Lemme, A., Ricque-Marie, D. (2009). Apparent drymatter, energy, protein and amino acid digestibility of four soybean ingredients in white shrimp *Litopenaeus vannamei* juveniles. Aquaculture, 292, 87–94. <https://doi.org/10.1016/j.aquaculture.2009.03.026>
8. Egamberdieva, D., Jabborova, D., Wirth, S.J., Alam, P., Alyemini, M.N., Ahmad, P. (2018). Interactive effects of nutrients and Bradyrhizobium japonicum on the growth and root architecture of soybean (*Glycine max* L.). Front. Microbiol., 9, 1–11. <https://doi.org/10.3389/fmicb.2018.01000>



**5. PENGIRIMAN REVISI ARTIKEL KE JOURNAL OF ECOLOGICAL ENGINEERING  
18 MEI 2023**



Muhibuddin &lt;muhibuddin@universitasbosowa.ac.id&gt;

---

**REVISION CONFIRMATION**

---

Muhibuddin <muhibuddin@universitasbosowa.ac.id>  
Kepada: office@jeeng.net

18 Mei 2023 pukul 11.48

Dear Editor Team,

We would like to thank you for your review of our article entitled: Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield. The manuscript has been revised and edited as reviewer's request.

Regards,  
Muhibuddin

---

 **Article Revision.doc**  
324K

## REVIEWER'S SPECIFIC COMMENTS PER SECTION OF MANUSCRIPT

	Reviewer	Author
Title	The title is mostly appropriate	
Abstract	The authors should correct all grammatical and typo	We apologise for the error. We have corrected the grammatical and typographical errors
Keywords	Keywords are appropriate	
Introduction	The authors should briefly discuss the relevance of the research methods to the research. Authors should correct all grammatical, spellings, italic and typo.	We have added relevant matters related to research methods to be researched. We have corrected the grammatical, spellings, italic and typographical errors. Hopefully this revised manuscript can be more comfortable to read.
Materials and Methods	The authors should improve the table to make it easy to read. Authors should clarified using what statistical analysis. All grammatical, spellings and typo should be corrected.	We have fixed all the existing tables. hopefully the tables can be read properly. We have described the statistical analysis used. Hopefully this revised manuscript can be more comfortable to read.
Results and Discussion	The authors should improve the table to make it easy to read. All grammatical, spellings and typo should be corrected.	We have fixed all the existing tables. We have corrected the grammatical and typographical errors
Conclusion	Conclusion are appropriate	
References	The authors should delete brackets in whole reference	We have removed all the brackets



# Utilization of Dry Land Using Molybdenum, Lime, and *Rhizobium* Strains to Increase Soybean Yield

Zulkifli Maulana<sup>1</sup>, Andi Muhibuddin<sup>1\*</sup>, Muhammad Arief Nasution<sup>1</sup>, Abri Abri<sup>1</sup>, Amiruddin Amirudin<sup>1</sup>, Baharuddin Baharuddin<sup>2</sup>, Andi Tenri Fitriyah<sup>2</sup>, Sitti Nurani Sirajuddin<sup>3</sup>, Abdel Razzaq M. Al Tawaha<sup>4</sup>

<sup>1</sup> Department of Agrotechnology, Faculty of Agriculture, Bosowa University, Makassar, 90245, Indonesia

<sup>2</sup> Department of Agribusiness, Faculty of Agriculture, Bosowa University Makassar, 90245, Indonesia

<sup>3</sup> Hasanuddin University, Socio-Economics Husbandry Department, Faculty of Animal Science, Makassar, Indonesia

<sup>4</sup> Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia 43400 UPM Serdang, Selangor, Malaysia

\* Corresponding author's e-mail: muhibuddin@universitasbosowa.ac.id

## ABSTRACT

Ultisol is a type of soil with low organic matter, pH, and nutrient content, including molybdenum, leading to low productivity. This study aimed to investigate the use of dry land using molybdenum and lime (CaCO<sub>3</sub>) inoculated with *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* to increase the soybean production of *Willis* and *Baluran* cultivars. This research was conducted from May to September 2021 in Pallangga Subdistrict, Gowa Regency, South Sulawesi, Indonesia. The study used a split-plot design with three replications for each treatment. The first factor was soybean varieties, consisting of *Baluran* and *Willis* cultivars. The second factor was the composition of the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>*, lime CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate, which consisted of without (*Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> + NH<sub>4</sub>-molybdate); *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 1.0 ton/ha + NH<sub>4</sub>-molybdate 250 g/h); *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/h); and *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/h). The results showed that treating the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>* + MoCo (1.0:0.6) kg/ha achieved the best results on growth, nutrient uptake (Nitrogen, Phosphorus and Potassium), and soybean yields, both for *Willis* and *Baluran* varieties on ultisol soils.

**Keywords:** Nod factor; nodulation; legume symbiosis; biological nitrogen fixation; rhizobium strain *nod<sup>+</sup>fix<sup>+</sup>*.

## INTRODUCTION

Indonesia's soybean demand continues to increase along with population growth. This is due to the need for soybeans in the community for consumption, so the increase in soybean production is one indicator of the success of the agricultural sector. The high demand for soybeans is because it is one of the basic needs and the main source of vegetable protein for most Indonesians. In addition, soybeans are a group of leguminous plants that can fix nitrogen from the air that occurs in root nodules (Al-Tawaha et al., 2022a; Al-Tawaha et al., 2022b; Saranraj et al., 2021; Shiri Janagard and Ebadi-Segherloo, 2015) with an average

protein content of 40% based on dry matter (Al-Tawaha and Al-Tawaha, 2017; Cruz-Suárez et al., 2009; Banaszekiewicz, 2011), soybean consumption in Indonesia is 2.2 million tons per year. This figure is far from the average production level of only 819,442 tons per year. As a result, the government must import soybeans as much as 2–2.6 million tons per year. Domestic soybean production is currently only able to meet about 35% of the consumption (Ningrum et al., 2018). The government has made several attempts to increase domestic soybean production, such as by clearing new farming land. Ultisol soil is one of the alternative lands for developing soybean areas in Indonesia, whose distribution reaches 45,794,000

ha or about 25% of the total land area of Indonesia (Syahputra et al., 2015), with great potential to be developed into productive land. However, ultisol soils are poor in organic matter (<5%), low pH, high Al saturation, low base saturation, and organic C, causing nutrient deficiencies, especially molybdenum (Mo) (Fitriatin et al., 2014). On the other hand, the element Mo is needed in the nitrate reductase enzyme to reduce nitrogen in nitrogen fixation in the root nodule tissue of soybean plants (Stiens et al., 2007). The element Mo is a component of the nitrate reductase enzyme, which plays a significant role in the assimilation of nitrogen nitrate (Mmbaga et al., 2015). High acidity (low pH) in ultisol soils and high Al saturation values can be neutralized by liming. Lime (CaCO<sub>3</sub>) increases pH, neutralizes Al and Fe, and increases soil Ca levels so that it can affect soil potassium levels to achieve balanced nutrient adequacy for soybean plants (Rahman et al., 2018). With a decrease in soil pH, *Rhizobium* bacteria can adapt, i.e., in the low pH range, around pH 5.0–6.5 (Cieřla et al., 2016). This is because root nodules of leguminous plants will only form when *Rhizobium* bacteria are present in the place where legumes grow (Wang et al., 2018). However, air nitrogen fixation does not always occur efficiently due to competition between inoculums which are efficient and inefficient specific *Rhizobium* that exist naturally in the soil (Mitsch et al., 2007; Mengel et al., 2012).

According to Reuhs et al. (2005), several types of *Rhizobium* bacteria are known based on their ability to form nodules, i.e., a) *Rhizobium* which does not have a nodule-forming character on plant roots, so that no symbiosis occurs, is known as a *Nod<sup>-</sup>* strain; b) *Rhizobium* which has nodule-forming character but does not have air nitrogen fixing character, known as *Nod<sup>+</sup>Fix<sup>-</sup>* strain; and c) *Rhizobium* which has two characters at once as nodule forming and as air nitrogen fixer is known as *Nod<sup>+</sup>Fix<sup>+</sup>* strain (Salih et al., 2015). Research on the inoculation of *Rhizobium* strains and the application of molybdenum to soybean plants has been carried out by several previous researchers (Lande et al., 2019). However, no research has combined the treatment of molybdenum, CaCO<sub>3</sub>, NH<sub>4</sub>-molybdate with *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* in ultisol soils.

The urgency of this research was the breakthrough of soybean production technology innovation by combining molybdenum, CaCO<sub>3</sub>, NH<sub>4</sub>-molybdate with *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*

on ultisol soils which have not been widely used in Indonesia because it is a marginal soil group. In addition, this research can reduce Indonesia's dependence on imported soybeans. Indonesia is currently still importing 2.6 million tons of soybeans from abroad.

This study aimed to investigate the use of dry land using molybdenum and CaCO<sub>3</sub> inoculated with *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* to increase the soybean production of *Willis* and *Baluran* cultivars.

## MATERIALS AND METHODS

### Experimental design and treatments

This research was conducted in the greenhouse of the Faculty of Agriculture, University of Bosowa, Pallangga Subdistrict, Gowa Regency, South Sulawesi, Indonesia, from May to September 2021. The study was arranged according to a split-plot design with two factors, with three replications for 24 plots, and each plot had 20 plants. Five plants were measured as the samples for each plot. The first factor was soybean varieties consisting of V<sub>1</sub> = *Baluran* and V<sub>2</sub> = *Willis*. The second factor was the composition of the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>*, lime CaCO<sub>3</sub> and NH<sub>4</sub>-molybdates follows: R<sub>0</sub> = without (*Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> + NH<sub>4</sub>-molybdate); R<sub>1</sub> = *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 24 g/pot + NH<sub>4</sub>-molybdate 6 g/pot; R<sub>2</sub> = *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 36 g/pot + NH<sub>4</sub>-molybdate 12 g/pot; dan R<sub>3</sub> = *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 48 g/pot + NH<sub>4</sub>-molybdate 18 g/pot.

### Preparation of growing media, administration of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*, and treatment of CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate

The soil used was ultisol soil which was taken compositely at a depth of 0–25 cm and had never been planted with soybeans. The soil was then air-dried and crushed and sieved with a 0.5 cm diameter sieve. The sieved soil was taken sufficiently to determine its physical and chemical properties in the laboratory before the treatments (Table 1). After that, 10 kg of ultisol soil was put into a black polybag with a size of 60 cm and a diameter of 40 cm and CaCO<sub>3</sub> was added (according to the treatment). The polybags were arranged in rows with wooden supports, with a distance between rows of 40 cm and between

**Table 1.** Chemical and physical properties of ultisol soils before and after treatments

Parameters	Content before treatment ( $R_0$ )	Content after treatment ( $R_1$ )	Content after treatment ( $R_2$ )	Content after treatment ( $R_3$ )
pH (1:2.5) H <sub>2</sub> O	4.9	5.50	5.65	5.70
C-organic (%)	0.82	0.83	0.83	0.84
N-Total (%)	0.18	0.28	0.36	0.38
Available-P (ppm)	10.00	10.20	10.37	10.48
K-dd (me/100 g)	0.32	0.43	0.47	0.51
CEC (me/100 g)	15.07	15.10	16.10	16.20
Base saturation (%)	15.15	16.10	16.12	16.40
Al-dd (me/100 g)	1.92	10.20	10.20	10.20
Mo (ppm)	0.35	1.51	1.69	2.10
Al saturation	22.64	22.12	21.78	21.60
Fraktion – sand (%)	55.23	55.21	55.21	55.10
Dust (%)	24.10	24.12	24.13	24.32
Clay (%)	20.67	20.67	20.66	20.58
Tekstur	Sandy clay loam	Sandy clay loam	Sandy clay loam	Sandy clay loam

rows of 25 cm. One week later, 50 g/pot. Then pots were given with basic fertilizers of Urea, TSP and KCl each dose of 0.25; 50 and 0.25 g/pot (50, 100 and 50 kg/ha). 7.5 mL *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* was mixed with 1 kg of soybean seeds. The mixture was then moistened with water, drained, mixed well, and stored in a shady place. It was left for 15 minutes and then was planted. The treatment of NH<sub>4</sub>-molybdate as a source of Mo (containing 82% acid MoO<sub>3</sub>) (according to the treatment) was given at the planting time by dissolving each in 10 ml of water. The solution was sprinkled into polybags around the plants and then sprayed on the plants once every two weeks. After one week, the plants were thinned so only one plant/polybag was maintained until the plants were harvested. After the plants were harvested, the soil residue from the polybags was taken compositely at a depth of 0–25 cm to analyze its physical and chemical properties in the laboratory.

### Maintenance and harvest

Watering was performed daily by maintaining the groundwater condition at around 80% of field capacity. Thinning was carried out after the plants were 7 days old by removing one plant per polybag. Plant maintenance included embroidery, weeding, and hoarding. Plants were harvested after about three months of age. The characteristics of soybean plants that were ready to harvest were that, after all, the plant leaves were old or yellow.

### Observation and measurement

Vegetative growth observation included plant height, age of flower appearance, number of root nodules, percentage of effective root nodules, and nitrogen (N) and potassium (K) uptake. Meanwhile, observations for the generative phase were the weight of 100 seeds, the number of pods per plant, and dry seed weight per plant.

### Data analysis

The data from the experiment were statistically analyzed using SPSS version 16 software. The differences among treatments were compared using analysis of variance (ANOVA) with the least significant difference (LSD) posthoc test at a 5% probability level.

## RESULTS AND DISCUSSION

### Soil chemical and physical properties

The soil analysis results before (without treatment) and after the experiment (soil treatment results of mixed  $R_1$ ,  $R_2$  and  $R_3$ ) showed a change in the chemical properties of the soil. In general, there was a change in the chemical properties of the soil, i.e., pH, organic matter, total nitrogen (N), available phosphorus (P), potassium (K), Cation-exchange capacity (CEC), Ca, Mg, Mo, and Al. This was due to the treatment of *Rhizobium* strain

*Nod<sup>+</sup>Fix<sup>+</sup>*, supplementation of microelements Mo (NH<sub>4</sub>-molybdate), and lime CaCO<sub>3</sub> in ultisol soils which contributed to improving the chemical properties of the soil. Application of NH<sub>4</sub>-molybdate increased Mo from 0.35 ppm to 0.47 ppm. The application of lime CaCO<sub>3</sub> increased soil pH from 5.1 to 5.5. This occurred due to the ability of lime CaCO<sub>3</sub> to neutralize soil acidity through the mechanism of H<sup>+</sup> substitution by Ca<sup>2+</sup>. This condition provides a way for the availability of phosphorus freely through mineralization by soil biota in the solution. The basic reason for liming is to increase soil pH. As a result of changes in pH, changes in soil chemistry and biology occur, such as increased availability of phosphorus, nitrogen, potassium, calcium, and magnesium.

### Vegetative growth

The ANOVA results showed that cultivars (V) and the combinations of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate (R) had a significant effect on plant height, age of flower appearance, and number of root nodules, and percentage of effective root nodules (Table 2). There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate. The *Baluran* cultivar (V1) produced higher plant height, age of flower emergence,

number of root nodules, and percentage of effective nodules. This was significantly different from the *Willis* cultivar (V2). These differences were due to the different characteristics of *Baluran* and *Willis* cultivars that affect photosynthetic activity, which determines plant height growth, flower appearance age, number of root nodules, and percentage of effective root nodules. Pregitzer et al. (2013) explained that some factors that influence plant growth and production are the environment, species or types of plants related to the genotype.

Table 2 shows that plant height, age of flower appearance, number of root nodules, and percentage of effective root nodules were increased with increasing doses of the combinations (CaCO<sub>3</sub>+NH<sub>4</sub>-molybdate). The highest increase was obtained in the combination of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*+ CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/ha of 90.16<sup>c</sup> cm, 35.66 days, 57.22 seeds, and 79.76 (%), respectively. However, the results for the combination of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*+ CaCO<sub>3</sub> 2.0 tons/ha+NH<sub>4</sub>-molybdate 750 g/ha were decreased statistically significantly. The increase in plant height, age of flower appearance, number of root nodules, and percentage of effective root nodules was supported by the availability of Mo for *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* by Mo administration. According to Schwarz and Mendel (2006), Mo is not only essential for plants, but also essential for microorganisms. Furthermore, more than 40 Mo enzymes are found in nature, most of which are found in bacteria, but only four Mo enzymes are found in plants. The increase in the availability of Mo in the soil has stimulated the growth, development, and activity of *Rhizobium* bacteria. Therefore, it increased the ability of *Rhizobium* bacteria to infect root nodules, resulting in a higher percentage of effective root nodules formed. Effective root nodules contain *Rhizobium* bacteria that can fix air N<sub>2</sub>.

Based on the results of Duncan's test in Table 2, the observation of plant height showed that the *Willis* cultivar achieved a higher plant height and was significantly different compared to the *Baluran* cultivar, both at the age of 2, 4, and 6 (weeks after planting) WAP, as well as significant differences in each treatment (R<sub>0</sub>, R<sub>1</sub>, R<sub>2</sub>, and R<sub>3</sub>). This was because the effectiveness of each treatment (bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>*, Lime (CaCO<sub>3</sub>), Molybdenum (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O) and Cobalt (CoC<sub>17</sub>)) also depends on the type of variety used.

**Table 2.** Average plant height, age of flower appearance, number of root nodules, and percentage of effective root nodules on soybean cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate

Treatment		Plant height (cm)	Age of flower appearance (days)	Number of root nodules (seeds)	Percentage of effective root nodules (%)
Cultivars	V1	83.96 <sup>a</sup>	32.78 <sup>a</sup>	40.22 <sup>a</sup>	62.21 <sup>a</sup>
	V2	75.62 <sup>b</sup>	38.96 <sup>b</sup>	34.64 <sup>b</sup>	58.28 <sup>b</sup>
Sub-plots	R0	66.04 <sup>a</sup>	35.38 <sup>a</sup>	09.80 <sup>a</sup>	27.23 <sup>a</sup>
	R1	82.59 <sup>b</sup>	36.05 <sup>a</sup>	26.14 <sup>b</sup>	67.76 <sup>b</sup>
	R2	90.16 <sup>c</sup>	35.66 <sup>a</sup>	57.22 <sup>cd</sup>	79.76 <sup>cd</sup>
	R3	80.37 <sup>b</sup>	36.10 <sup>a</sup>	48.47 <sup>c</sup>	73.74 <sup>c</sup>

**Note:** Numbers followed by the same letter in each column are not significantly different at Duncan's test level =0.05.

One factor affecting growth and production is the species or types of plants related to genotype (Pregitzer et al., 2013).

The results of Duncan's test in Table 2 on the observation of the number of root nodules showed that different treatments gave different results on the number of root nodules. Treatment R<sub>2</sub> showed the highest number of root nodules. This was due to the suitability of the needs for bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>*, Lime (CaCO<sub>3</sub>), Molybdenum (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O, and Cobalt (CoC<sub>12</sub>). Therefore, metabolic processes were increased due to increased assimilate formation, the influence of microelements Molybdenum and Cobalt, which play an essential role in enzyme systems (Bhattacharya et al., 2016). However, the soil analysis results before the study showed that Mo levels were quite low (0.97 ppm. In contrast, the requirement for soybeans was generally around 0.51 - 1.00 ppm (Ibañez et al., 2020). If the concentration of Mo in the soil is low, it will interfere with the formation of root nodules and inhibit N fixation by *Rhizobium* (Egamberdieva et al., 2018). This can be observed from the ability of plants to form effective root nodules. Ismunadji and Mahmud (1993) stated that applying Mo to soybean plants would increase the number of root nodules.

The increasing in the number of root nodules was caused by an increase in the number of nitrogenation enzymes and nitrate reductase, thereby increasing N fixation. (Imran et al., 2019) stated that Mo is the main component of the nitrogenation enzyme and nitrate reductase, whose preparation mechanism occurs due to a change in valence. Meanwhile, the process of forming root nodules occurs through cooperation between plants and N-fixing bacteria. Nodules that are active in symbiotic N binding, when split, will turn red due to the presence of leghaemoglobin. If the center of the root nodule is not red, it means it is not effective. Furthermore, (Zheng et al., 2020) and (Koskey et al., 2018) explained that root nodules are distinguished into effective and ineffective root nodules, in which the differences between the two root nodules are in size or shape, color, and location on the root.

The results of Duncan's test in Table 2 showed that the R<sub>2</sub> treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* + MoCo (1.0 : 0.6) kg/ha) produced the highest number of red nodules compared to other treatments. The high results indicated that the R<sub>2</sub> treatment impacted the formation of red root nodules. Therefore, it can infect plant roots in the

form of root nodules which are effective in fixing nitrogen from the air. This is presumably due to the ability of inoculation to meet the optimal N requirement for plant growth forming an effective red nodule. In addition, root nodules that are effective at fixing N can be recognized by their reddish color on the inside when the nodules are split. The red color is due to the presence of the pigment leghaemoglobin.

The observation results of the distribution pattern of the root nodules in Table 2 showed the best treatment of R<sub>2</sub>. The nodule distribution pattern determines the effectiveness of the nodule. Nodules scattered on the root neck indicate nodules effective in fixing N. The distribution pattern of root nodules explains the effectiveness of root nodules in fixing nitrogen free from the air. Effective root nodules are located on the taproot, while ineffective root nodules are scattered on lateral roots or root branches and are pale in color. The activity of N<sub>2</sub> fixation by bacteria is essential for overall N balance because the continuous form of fixed N can be lost due to denitrification and permeation (Howie & Echandi, 1983).

### Nitrogen, phosphorus and potassium uptake

The ANOVA results showed that cultivars (V) had a significant effect on nitrogen (N) and potassium (K) uptake but had no significant effect on Phosphorus (P) uptake. The combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate (R) significantly affected N, P, and K uptake (Table 3). There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate.

The *Baluran* cultivar (V<sub>1</sub>) produced higher N and K uptake, significantly different from the *Willis* cultivar (V<sub>2</sub>). However, the P uptake of treatments was not significantly different. According to

Table 3 shows that N, P, and K uptake increased with increasing doses of the combinations (CaCO<sub>3</sub> + NH<sub>4</sub>-molybdate). The highest increase was obtained in the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/ha of 852.7 mg/plant, 52.4 mg/plant, and 442.9 mg/plant, respectively. However, the results for the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/ha were decreased statistically significantly. The increased uptake of N, P, and K was thought to be the result of the ability of soybean cultivars to form root nodules in

**Table 3.** Average nutrient uptake of N, P and K in soybean cultivars and combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate

Treatment		Nutrient uptake (mg/plant)		
		N	P	K
Cultivars	V1	710.6 <sup>a</sup>	43.1 <sup>a</sup>	370.0 <sup>a</sup>
	V2	641.1 <sup>a</sup>	40.5 <sup>a</sup>	364.7 <sup>a</sup>
Sub-plots	R0	385.6 <sup>a</sup>	33.1 <sup>a</sup>	244.8 <sup>a</sup>
	R1	737.3 <sup>b</sup>	41.2 <sup>b</sup>	377.3 <sup>b</sup>
	R2	852.7 <sup>c</sup>	52.4 <sup>c</sup>	442.9 <sup>bc</sup>
	R3	727.8 <sup>b</sup>	40.6 <sup>b</sup>	405.4 <sup>b</sup>

**Note:** numbers followed by the same letter in each column are not significantly different at Duncan's test level = 0.05.

response to *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* inoculation. In addition, it was supposed that the number of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* in that treatment was more suitable than others. The root nodules were formed if there was a suitability between *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* and its host.

The results of Duncan's test in Table 3 showed that the highest nitrogen uptake was found in the R<sub>2</sub> treatment compared to other treatments (R<sub>0</sub>, R<sub>1</sub> and R<sub>3</sub>). The significant effect of the R<sub>2</sub> treatment (bacterial strain *Nod Fix* + MoCo (1.0 : 0.6) kg/ha) was thought to result from the soybean's ability to form root nodules in response to the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* inoculation. In addition, it is suspected that the number of bacterial strains *Nod<sup>+</sup> Fix<sup>+</sup>* in this treatment is more suitable than the other treatments, and effective root nodules would form if there were a suitability between the bacteria and its host.

### Leaf color scale

The observation results of the leaf color scale in Table 4 showed that the treatment of bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* with Mo and CaCO<sub>3</sub> lime supplements on soybeans using ultisol soils affected the leaf color scale. Table 4 shows that the *Willis* and *Baluran* varieties treated with the *Nod<sup>+</sup> Fix<sup>+</sup>* bacterial strain, Mo and CaCO<sub>3</sub> lime supplements had various leaf color scales.

Table 4 showed that the results on the leaf color scale was the best treatment of R<sub>1</sub> for the *Baluran* cultivar compared to other treatments. This showed that the fixed N element was used to meet the N required by plants so the availability of N for plants increases. An effective fixation, 50–75% of the total N requirement by plants can be fulfilled. In general, this study showed the pattern of increasing plant height, root dry weight, number of root nodules, number of red nodules,

**Table 4.** Leaf color scale

Varietas	Treatments	Leaf color scales *			
		1	2	3	4
V <sub>1</sub>	R <sub>0</sub>	x			
	R <sub>1</sub>		x		
	R <sub>2</sub>			x	
	R <sub>3</sub>			x	
V <sub>2</sub>	R <sub>0</sub>	x			
	R <sub>1</sub>				x
	R <sub>2</sub>			x	
	R <sub>3</sub>			x	

**Note:** \* Criteria based on color scale, diagnosis of nutrient status from leaf color. The strengthening of legumes in relation to cropping system research project. Japan International Cooperation Agency (JICA): 1 – yellowish green, 2 – light green, 3 – green, 4 – slightly dark green.

N uptake, root nodule distribution pattern, leaf color scale, weight of 100 seeds, and dry seed yield. It was found that the treatment of the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* with supplements of microelements (MoCo), lime (CaCO<sub>3</sub>), and Cobalt, simultaneously increased the availability of Mo in the soil so as to increase the uptake of Mo by plants. Jaiswal et al. (2021) stated that the element Mo functions as an activator of nitrogenation and reducing enzymes which activate *Rhizobium* performance in the N fixation process so that N binding increases. Nitrogen can stimulate cell elongation, vegetative growth, enlarge pods, slow down the initiation phase, and increase seeds yield and content (Ohyama et al., 2013). The results of Duncan's test in Table 4 showed that the highest weight of 100 seeds was obtained in the R<sub>2</sub> treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* + MoCo (1.0 : 0.6) kg/ha) compared to other treatments (R<sub>0</sub>, R<sub>1</sub> and R<sub>3</sub>). This revealed that the treatment is vital for the growth and production of soybeans.

**Table 5.** The average weight of 100 seeds, number of pods per plant, and dry seed weight per plant

Treatment		Weight 100 seeds (g)	Number of pods per plant (piece)	Dry seed weight per plant (g)
Cultivars	V1	13.75 <sup>a</sup>	28.66 <sup>a</sup>	16.53 <sup>a</sup>
	V2	11.34 <sup>a</sup>	26.51 <sup>a</sup>	15.17 <sup>a</sup>
Sub-plots	R0	10.37 <sup>a</sup>	19.18 <sup>a</sup>	10.99 <sup>a</sup>
	R1	14.04 <sup>b</sup>	26.94 <sup>b</sup>	17.97 <sup>bc</sup>
	R2	14.49 <sup>b</sup>	35.89 <sup>c</sup>	19.45 <sup>c</sup>
	R3	11.88 <sup>a</sup>	28.34 <sup>b</sup>	15.00 <sup>b</sup>

Note: numbers followed by the same letter in each column are not significantly different at Duncan's test level = 0.05.

This was due to the Cofactor Mo (MoCo) forming the active site of all eukaryotic molybdenum enzymes.

### Yield components

The ANOVA results showed that cultivars (V) had no significant effect; while the combinations of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate had a significant effect on the weight of 100 seeds, number of pods per plant, and dry seed weight per plant. There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate (Table 5).

Table 5 shows that the weight of 100 seeds, number of pods per plant, and dry seed weight per plant were increased with increasing doses of the combinations (CaCO<sub>3</sub>+ NH<sub>4</sub>-molybdate), the highest increase was achieved in the combination of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*+ CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/ha of 14.49 g, 35.89 pieces, and 19.45 g, respectively. However, the results for the combination of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/ha decreased statistically significantly. Increased vegetative growth continues in the generative growth of plants. The excess photosynthate, as a result of the photosynthesis process, was utilized to form pods, then stored in the form of seeds. In addition, the increase in N produced by plants will increase the synthesis of amino acids and proteins and plant organic compounds. These organic compounds are deposited on the seeds, increasing the dry seed weight per plant.

At the combination dose of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/ha, there was a tendency to decrease growth and yield of soybeans, presumably due to the excess of Mo and CaCO<sub>3</sub>. Based on Table

5, the highest dry seed yield was obtained in treatment R<sub>2</sub> and was significantly different compared to other treatments (R<sub>0</sub> and R<sub>3</sub>). This was because the R<sub>2</sub> treatment will increase nitrogen fixation to meet nitrogen needs for plants so that the availability of nitrogen for plants increases. In the R<sub>3</sub> treatment, there was a tendency to decrease the growth and production of soybeans, presumably due to excess (toxicity) of Mo and Co. The elements Mo and Co are micronutrients that are required in relatively low amounts with a narrow optimum range (Maggini et al., 2018). Therefore, its slightly excessive amounts will interfere with plant physiological processes and can even be toxic to plants. According to (Glass et al., 2012), the addition of excessive Mo will not affect the fixation of N.

The results of soil analysis before and after the study showed a change/increase in the content of physical/chemical properties of the soil, including parameters of soil pH, organic matter; N-total; Phosphor; Potassium; CEC; Ca; Mg; Mo and Co. This was due to the administration of the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>* with Micro (MoCo) and Lime (CaCO<sub>3</sub>) supplementations. In the podzolic marginal soil, the chemical properties were generally low/moderate (Table 1), and in each of these treatments, the *Nod<sup>+</sup>Fix<sup>+</sup>* strain bacteria, Lime (CaCO<sub>3</sub>), Molybdenum (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O, and Cobalt (CoC<sub>12</sub>) contributed to improving the chemical properties of the podzolic soil.

### CONCLUSION

This study showed that treating the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>*+ MoCo (1.0 : 0.6) kg/ ha gave the best results on soybean growth and production, both for *Willis* and *Baluran* varieties on Ultisol marginal soils. Increasing the Mo dose will not significantly increase plant growth and yield.

## Acknowledgments

This work was supported by providing facilities from the Bosowa University, Indonesia.

## REFERENCES

1. Al-Tawaha, A.R.M., McNeil, D., Othman, Y.A., AlRawashdeh, I.M., Adnan, M., Zamin, M., Qaisi, A.M., Al-Tawaha, A., Jahan, N., Shah, M.A., Khalid, S. 2022a. Legume Production and Climate Change. In *Improvement of Plant Production in the Era of Climate Change*. CRC Press, 221–248.
2. Al-Tawaha, A.R.M.S., Khanum, S., Benkeblia, N., Khalid, S., Al-Tawaha, A.R., Mondal, M., Odat, N., Dey, A., Alimad, N., Thangadurai, D., Sangeetha, J. 2022b. *Adapting Crops to Climate Change. Climate Change and Agriculture: Perspectives, Sustainability and Resilience*, 53–77. <https://doi.org/10.1002/9781119789789.ch3>
3. Al-Tawaha, A.R., Al-Tawaha, A.R.M. 2017. Response of soybean plants to exogenous application of yeast extract: Growth and chemical composition. *American-Eurasian Journal of Sustainable Agriculture*, 11(2), 31–36.
4. Banaszkiwicz, T. 2011. Nutritional value of soybean meal. *Soybean and Nutrition*, 1–21. <https://doi.org/10.5772/23306>
5. Bhattacharya, P.T., Misra, S.R., Hussain, M. 2016. Nutritional aspects of essential trace elements in oral health and disease: an extensive review. *Scientifica*, 1–12. <https://doi.org/10.1155/2016/5464373>
6. Cieřla, J., Kopycińska, M., Łukowska, M., Bieganski, A., Janczarek, M. 2016. Surface properties of wild-type *Rhizobium leguminosarum* bv. *Trifolii* strain 24.2 and Its derivatives with different extracellular polysaccharide content. *PLoS ONE*, 11(10), 1–21. <https://doi.org/10.1371/journal.pone.0165080>
7. Cruz-Suárez, L.E., Tapia-Salazar, M., Villarreal-Cavazos, D., Beltran-Rocha, J., Nieto-López, M.G., Lemme, A., Ricque-Marie, D. 2009. Apparent dry matter, energy, protein and amino acid digestibility of four soybean ingredients in white shrimp *Litopenaeus vannamei* juveniles. *Aquaculture*, 292, 87–94. <https://doi.org/10.1016/j.aquaculture.2009.03.026>
8. Egamberdieva, D., Jabborova, D., Wirth, S.J., Alam, P., Alyemeni, M.N., Ahmad, P. 2018. Interactive effects of nutrients and Bradyrhizobium japonicum on the growth and root architecture of soybean (*Glycine max* L.). *Front.Microbiol.*, 9, 1–11. <https://doi.org/10.3389/fmicb.2018.01000>
9. Fitriatin, B.N., Yuniarti, A., Turmuktini, T., Ruswandi, F.K. 2014. The effect of phosphate solubilizing microbe producing growth regulators on soil phosphate, growth and yield of maize and fertilizer efficiency on Ultisol. *Eurasian J. Soil Sci.*, 3(2), 101–107. <https://doi.org/10.18393/ejss.34313>
10. Glass, J.B., Axler, R.P., Chandra, S., Goldman, C.R. 2012. Molybdenum limitation of microbial nitrogen assimilation in aquatic ecosystems and pure cultures. *Front. Microbiol.*, 3, 1–11. <https://doi.org/10.3389/fmicb.2012.00331>
11. Howie, W.J., Echandi, E. 1983. Rhizobacteria: influence type on plant growth and yield of potato. *Soil Biol. Biochem.*, 15(2), 127–132.
12. Ibañez, T.B., Santos, L.F. de M., Lapaz, A. de M., Ribeiro, I.V., Ribeiro, F.V., Reis, A.R. Dos, Moreira, A., Heinrichs, R. 2020. Sulfur modulates yield and storage proteins in soybean grains. *Sci. Agric.*, 78(1), 1–9. <https://doi.org/10.1590/1678-992x-2019-0020>
13. Imran, M., Sun, X., Hussain, S., Ali, U., Rana, M.S., Rasul, F., Saleem, M.H., Moussa, M.G., Bhantana, P., Afzal, J., Elyamine, A.M., Hu, C.X. 2019. Molybdenum-induced effects on nitrogen metabolism enzymes and elemental profile of winter wheat (*Triticum aestivum* L.) under different nitrogen sources. *Int. J. Mol. Sci.*, 20(3009), 1–17. <https://doi.org/10.3390/ijms20123009>
14. Jaiswal, S.K., Mohammed, M., Ibny, F.Y.I., Dakora, F.D. 2021. Rhizobia as a source of plant growth-promoting molecules: potential applications and possible operational mechanisms. *Front. Sustain. Food Syst.*, 4, 1–14. <https://doi.org/10.3389/fsufs.2020.619676>
15. Koskey, G., Mburu, S.W., Kimiti, J.M., Ombori, O., Maingi, J.M., Njeru, E.M. 2018. Genetic characterization and diversity of Rhizobium isolated from root nodules of mid-altitude climbing bean (*Phaseolus vulgaris* L.) varieties. *Front. Microbiol.*, 9, 1–12. <https://doi.org/10.3389/fmicb.2018.00968>
16. Lande, M., Bawankule, K.V., Solanki, R.D., Aware, R.G. 2019. Effect of bradyrhizobium broth on growth of root, shoot and nodule of soybean. *Int.J. Curr. Microbiol. Appl. Sci.*, 8(7), 1588–1596. <https://doi.org/10.20546/ijcmas.2019.807.189>
17. Maggini, S., Pierre, A., Calder, P.C. 2018. Immune function and micronutrient requirements change over the life course. *Nutrients*, 10(1531), 1–27. <https://doi.org/10.3390/nu10101531>
18. Mengel, D., Ruiz-diaz, D., Asebedo, R., Maxwell, T. 2012. Nitrogen fertilization of nitrogen-stressed soybeans. *Better Crops*, 96, 14–15.
19. Mitsch, M.J., Cowie, A., Finan, T.M. 2007. Malic enzyme cofactor and domain requirements for symbiotic N<sub>2</sub> fixation by *Sinorhizobium meliloti*. *J. Bacteriol.*, 189, 160–168. <https://doi.org/10.1128/JB.01425-06>
20. Mmbaga, G.W.M., Mtei, K., Ndakidemi, P.A. 2015. Yield and fiscal benefits of inoculation supplemented with phosphorus (P) and Potassium (K) in Climbing Beans Grown in Northern Tanzania. *Agri. Sci.*,



- 6, 783–797. <https://doi.org/10.4236/as.2015.68076>
21. Ningrum, I.H., Irianto, H., Riptanti, E.W. 2018. Analysis of soybean production and import trends and its import factors in Indonesia. IOP Conference Series: Earth and Environmental Sci., 142, 1–8. <https://doi.org/10.1088/1755-1315/142/1/012059>
22. Ohyama, T., Minagawa, R., Ishikawa, S., Yamamoto, M., Phi Hung, N., Van, Ohtake, N., Sueyoshi, K., Sato, T., Nagumo, Y., Takahashi, Y. 2013. Soybean Seed Production and Nitrogen Nutrition. A Comprehensive Survey of International Soybean Research - Genetics, Physiology, Agronomy and Nitrogen Relationships, 115–157. <https://doi.org/10.5772/52287>
23. Pregitzer, C.C., Bailey, J.K., Schweitzer, J.A. 2013. Genetic by environment interactions affect plant-soil linkages. Ecol. Evol. 3, 2322–2333. <https://doi.org/10.1002/ece3.618>
24. Rahman, M.A., Lee, S.H., Ji, H.C., Kabir, A.H., Jones, C.S., Lee, K.W. 2018. Importance of mineral nutrition for mitigating aluminum toxicity in plants on acidic soils: Current status and opportunities. International Journal of Molecular Sciences, 79(3073), 1–18. <https://doi.org/10.3390/ijms19103073>
25. Reuhs, B.L., Relić, B., Forsberg, L.S., Marie, C., Ojanen-Reuhs, T., Stephens, S.B., Wong, C.H., Jabbouri, S., Broughton, W.J. 2005. Structural characterization of a flavonoid-inducible *Pseudomonas aeruginosa* A-band-like O antigen of *Rhizobium* sp. strain NGR234, required for the formation of nitrogen-fixing nodules. J. Bacteriol., 187, 6479–6487. <https://doi.org/10.1128/JB.187.18.6479-6487.2005>
26. Salih, S., Hamd, S.A.M., Dagash, Y.M.I. 2015. The effects of rhizobium, mycorrhizal inoculations and diammonium phosphate (DAP) on nodulation, growth, and yield of soybean. Univers J. Agri.Res., 3, 11–14. <https://doi.org/10.13189/ujar.2015.030103>
27. Shiri Janagard, M., Ebadi-Segherloo, A. 2015. Inoculated soybean response to starter nitrogen in conventional cropping system in Moghan. J. Agron., 15, 26–32. <https://doi.org/10.3923/ja.2016.26.32>
28. Stiens, M., Schneiker, S., Pühler, A., Schlüter, A. 2007. Sequence analysis of the 181-kb accessory plasmid pSmeSM11b, isolated from a dominant *Sinorhizobium meliloti* strain identified during a long-term field release experiment. FEMS Microbio. Lett., 271, 297–309. <https://doi.org/10.1111/j.1574-6968.2007.00731.x>
29. Syahputra, E., Fauzi, Razali. 2015. The characteristics of the chemical properties of ultisols sub groups in some areas of Northern Sumatra. Theor. Appl. Climatol., 4, 1796–1803. <http://dx.doi.org/10.1016/j.ecolecon.2013.05.006>
30. Saranraj, P., Sivasakthivelan, P., Al-Tawaha, A.R.M., Bright, R., Al-Tawaha, A.R., Thangadurai, D., Sangeetha, J., Rauf, A., Khalid, S., Al Sultan, W., Safari, Z.S. 2021. Macronutrient management for the cultivation of Soybean (*Glycine max* L.): A review. In IOP Conference Series: Earth and Environmental Science. IOP Publishing, 788(1), 012055.
31. Wang, Q., Liu, J., Zhu, H. 2018. Genetic and molecular mechanisms underlying symbiotic specificity in legume-rhizobium interactions. Front. Plant Sci., 9, 1–8. <https://doi.org/10.3389/fpls.2018.00313>
32. Zheng, Y., Liang, J., Zhao, D.L., Meng, C., Xu, Z.C., Xie, Z.H., Zhang, C.S. 2020. The root nodule microbiome of cultivated and wild halophytic legumes showed similar diversity but distinct community structure in yellow river delta saline soils. Microorganisms, 8, 1–12. <https://doi.org/10.3390/microorganisms8020207>

**6. KONFIRMASI PENERIMAAN REVISI PENULIS KE EDITOR**  
**19 MEI 2023**



Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

---

**New revision received by Editorial Office (JEENG-04241-2023-02)**

---

Journal of Ecological Engineering <office@jeeng.net>

19 Mei 2023 pukul 03.12

Kepada: muhibuddin <muhibuddin@universitasbosowa.ac.id>, Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

Dear Authors,

Thank you for the revision manuscript, entitled: Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield. The following number has been assigned to it JEENG-04241-2023-02.

The manuscript will be rated once again by the Editors and then sent to the Reviewers.

You will be informed by email about any further decision on this article.

Kindest regards,  
Justyna Kujawska  
Managing Editor  
Journal of Ecological Engineering

**7. KONFIRMASI PENERIMAAN ARTIKEL**  
**29 MEI 2023**



Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

---

## Decision on manuscript JEENG-04241-2023-02

---

Journal of Ecological Engineering <office@jeeng.net>

29 Mei 2023 pukul 06.11

Kepada: muhibuddin <muhibuddin@universitasbosowa.ac.id>, Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

May 29, 2023

JEENG-04241-2023-02

Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield

Dear Authors,

I am pleased to inform you that your manuscript, entitled: Utilization of Dry Land using Molybdenum, Lime, and Rhizobium Strains to Increase Soybean Yield, has been accepted with revision references, a few typo and others details.

Thank you for submitting your work to us.

Kindest regards,  
Justyna Kujawska  
Managing Editor  
Journal of Ecological Engineering

**8. KONFIRMASI PROOF VERSION**  
**19 JUNI 2023**



Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

---

## JEE 24(8), 2023

Journal of Ecological Engineering <office@jeeng.net>

19 Juni 2023 pukul 15.07

Kepada: muhibuddin <muhibuddin@universitasbosowa.ac.id>, Zulkifli Maulana <zulkifli.maulana@universitasbosowa.ac.id>

Dear Authors,

I am sending a proof version of the article for publication in the journal of Ecological Engineering, Vol. 24, Iss. 8, 2023.

Please read the final version of the work, and use the attached PDF file if you need to add your comments. Please send your acceptance to 23th of June.

Best regards,  
Rozalia Skiba  
Editorial Assistant  
Journal of Ecological Engineering

---

 **Article.pdf**  
512K

---

**Muhibuddin** <muhibuddin@universitasbosowa.ac.id>

Kepada: Journal of Ecological Engineering <office@jeeng.net>

I approve the final version of the article

[Kutipan teks disembunyikan]

## Utilization of Dry Land Using Molybdenum, Lime, and *Rhizobium* Strains to Increase Soybean Yield

Zulkifli Maulana<sup>1</sup>, Andi Muhibuddin<sup>1\*</sup>, Muhammad Arief Nasution<sup>1</sup>, Abri Abri<sup>1</sup>, Amiruddin Amirudin<sup>1</sup>, Baharuddin Baharuddin<sup>2</sup>, Andi Tenri Fitriyah<sup>2</sup>, Sitti Nurani Sirajuddin<sup>3</sup>, Abdel Razzaq M. Al Tawaha<sup>4</sup>

<sup>1</sup> Department of Agrotechnology, Faculty of Agriculture, Bosowa University, Makassar, 90245, Indonesia

<sup>2</sup> Department of Agribusiness, Faculty of Agriculture, Bosowa University Makassar, 90245, Indonesia

<sup>3</sup> Hasanuddin University, Socio-Economics Husbandry Department, Faculty of Animal Science, Makassar, Indonesia

<sup>4</sup> Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia 43400 UPM Serdang, Selangor, Malaysia

\* Corresponding author's e-mail: muhibuddin@universitasbosowa.ac.id

### ABSTRACT

Ultisol is a type of soil with low organic matter, pH, and nutrient content, including molybdenum, leading to low productivity. This study aimed to investigate the use of dry land using molybdenum and lime ( $\text{CaCO}_3$ ) inoculated with *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* to increase the soybean production of *Willis* and *Baluran* cultivars. This research was conducted from May to September 2021 in Pallangga Subdistrict, Gowa Regency, South Sulawesi, Indonesia. The study used a split-plot design with three replications for each treatment. The first factor was soybean varieties, consisting of *Baluran* and *Willis* cultivars. The second factor was the composition of the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>*, lime  $\text{CaCO}_3$  and  $\text{NH}_4$ -molybdate, which consisted of without (*Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* +  $\text{CaCO}_3$  +  $\text{NH}_4$ -molybdate); *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* +  $\text{CaCO}_3$  1.0 ton/ha +  $\text{NH}_4$ -molybdate 250 g/h); *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* +  $\text{CaCO}_3$  1.5 tons/ha +  $\text{NH}_4$ -molybdate 500 g/h); and *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* +  $\text{CaCO}_3$  2.0 tons/ha +  $\text{NH}_4$ -molybdate 750 g/h). The results showed that treating the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>* + MoCo (1.0:0.6) kg/ha achieved the best results on growth, nutrient uptake (Nitrogen, Phosphorus and Potassium), and soybean yields, both for *Willis* and *Baluran* varieties on ultisol soils.

**Keywords:** Nod factor; nodulation; legume symbiosis; biological nitrogen fixation; rhizobium strain *nod<sup>+</sup>fix<sup>+</sup>*.

### INTRODUCTION

Indonesia's soybean demand continues to increase along with population growth. This is due to the need for soybeans in the community for consumption, so the increase in soybean production is one indicator of the success of the agricultural sector. The high demand for soybeans is because it is one of the basic needs and the main source of vegetable protein for most Indonesians. In addition, soybeans are a group of leguminous plants that can fix nitrogen from the air that occurs in root nodules (Al-Tawaha et al., 2022a; Al-Tawaha et al., 2022b; Saranraj et al., 2021; Shiri Janagard and Ebadi-Segherloo, 2015) with an average

protein content of 40% based on dry matter (Al-Tawaha and Al-Tawaha, 2017; Cruz-Suárez et al., 2009; Banaszkievicz, 2011), soybean consumption in Indonesia is 2.2 million tons per year. This figure is far from the average production level of only 819,442 tons per year. As a result, the government must import soybeans as much as 2–2.6 million tons per year. Domestic soybean production is currently only able to meet about 35% of the consumption (Ningrum et al., 2018). The government has made several attempts to increase domestic soybean production, such as by clearing new farming land. Ultisol soil is one of the alternative lands for developing soybean areas in Indonesia, whose distribution reaches 45,794,000



ha or about 25% of the total land area of Indonesia (Syahputra et al., 2015), with great potential to be developed into productive land. However, ultisol soils are poor in organic matter (<5%), low pH, high Al saturation, low base saturation, and organic C, causing nutrient deficiencies, especially molybdenum (Mo) (Fitriatin et al., 2014). On the other hand, the element Mo is needed in the nitrate reductase enzyme to reduce nitrogen in nitrogen fixation in the root nodule tissue of soybean plants (Stiens et al., 2007). The element Mo is a component of the nitrate reductase enzyme, which plays a significant role in the assimilation of nitrogen nitrate (Mmbaga et al., 2015). High acidity (low pH) in ultisol soils and high Al saturation values can be neutralized by liming. Lime ( $\text{CaCO}_3$ ) increases pH, neutralizes Al and Fe, and increases soil Ca levels so that it can affect soil potassium levels to achieve balanced nutrient adequacy for soybean plants (Rahman et al., 2018). With a decrease in soil pH, *Rhizobium* bacteria can adapt, i.e., in the low pH range, around pH 5.0–6.5 (Cieřla et al., 2016). This is because root nodules of leguminous plants will only form when *Rhizobium* bacteria are present in the place where legumes grow (Wang et al., 2018). However, air nitrogen fixation does not always occur efficiently due to competition between inoculums which are efficient and inefficient specific *Rhizobium* that exist naturally in the soil (Mitsch et al., 2007; Mengel et al., 2012).

According to Reuhs et al. (2005), several types of *Rhizobium* bacteria are known based on their ability to form nodules, i.e., a) *Rhizobium* which does not have a nodule-forming character on plant roots, so that no symbiosis occurs, is known as a *Nod* strain; b) *Rhizobium* which has nodule-forming character but does not have air nitrogen fixing character, known as *Nod<sup>+</sup>Fix<sup>-</sup>* strain; and c) *Rhizobium* which has two characters at once as nodule forming and as air nitrogen fixer is known as *Nod<sup>+</sup>Fix<sup>+</sup>* strain (Salih et al., 2015). Research on the inoculation of *Rhizobium* strains and the application of molybdenum to soybean plants has been carried out by several previous researchers (Lande et al., 2019). However, no research has combined the treatment of molybdenum,  $\text{CaCO}_3$ ,  $\text{NH}_4$ -molybdate with *Rhizobium* Strain *Nod<sup>+</sup>Fix<sup>+</sup>* in ultisol soils.

The urgency of this research was the breakthrough of soybean production technology innovation by combining molybdenum,  $\text{CaCO}_3$ ,  $\text{NH}_4$ -molybdate with *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*

on ultisol soils which have not been widely used in Indonesia because it is a marginal soil group. In addition, this research can reduce Indonesia's dependence on imported soybeans. Indonesia is currently still importing 2.6 million tons of soybeans from abroad.

This study aimed to investigate the use of dry land using molybdenum and  $\text{CaCO}_3$  inoculated with *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>* to increase the soybean production of *Willis* and *Baturan* cultivars.

## MATERIALS AND METHODS

### Experimental design and treatments

This research was conducted in the greenhouse of the Faculty of Agriculture, University of Bosowa, Pallangga Subdistrict, Gowa Regency, South Sulawesi, Indonesia, from May to September 2021. The study was arranged according to a split-plot design with two factors, with three replications for 24 plots, and each plot had 20 plants. Five plants were measured as the samples for each plot. The first factor was soybean varieties consisting of  $V_1 = \text{Baturan}$  and  $V_2 = \text{Willis}$ . The second factor was the composition of the bacterial strain *Nod<sup>+</sup>Fix<sup>+</sup>*, lime  $\text{CaCO}_3$  and  $\text{NH}_4$ -molybdates follows:  $R_0 = \text{without (Rhizobium strain Nod<sup>+</sup>Fix<sup>+</sup> + CaCO}_3 + \text{NH}_4\text{-molybdate)}$ ;  $R_1 = \text{Rhizobium strain Nod<sup>+</sup>Fix<sup>+</sup> + CaCO}_3 \text{ 24 g/pot + NH}_4\text{-molybdate 6 g/pot}$ ;  $R_2 = \text{Rhizobium strain Nod<sup>+</sup>Fix<sup>+</sup> + CaCO}_3 \text{ 36 g/pot + NH}_4\text{-molybdate 12 g/pot}$ ; dan  $R_3 = \text{Rhizobium strain Nod<sup>+</sup>Fix<sup>+</sup> + CaCO}_3 \text{ 48 g/pot + NH}_4\text{-molybdate 18 g/pot}$ .

### Preparation of growing media, administration of *Rhizobium* strain *Nod<sup>+</sup>Fix<sup>+</sup>*, and treatment of $\text{CaCO}_3$ and $\text{NH}_4$ -molybdate

The soil used was ultisol soil which was taken compositely at a depth of 0–25 cm and had never been planted with soybeans. The soil was then air-dried and crushed and sieved with a 0.5 cm diameter sieve. The sieved soil was taken sufficiently to determine its physical and chemical properties in the laboratory before the treatments (Table 1). After that, 10 kg of ultisol soil was put into a black polybag with a size of 60 cm and a diameter of 40 cm and  $\text{CaCO}_3$  was added (according to the treatment). The polybags were arranged in rows with wooden supports, with a distance between rows of 40 cm and between

**Table 1.** Chemical and physical properties of ultisol soils before and after treatments

Parameters	Content before treatment ( $R_0$ )	Content after treatment ( $R_1$ )	Content after treatment ( $R_2$ )	Content after treatment ( $R_3$ )
pH (1:2.5) $H_2O$	4.9	5.50	5.65	5.70
C-organic (%)	0.82	0.83	0.83	0.84
N-Total (%)	0.18	0.28	0.36	0.38
Available-P (ppm)	10.00	10.20	10.37	10.48
K-dd (me/100 g)	0.32	0.43	0.47	0.51
CEC (me/100 g)	15.07	15.10	16.10	16.20
Base saturation (%)	15.15	16.10	16.12	16.40
Al-dd (me/100 g)	1.92	10.20	10.20	10.20
Mo (ppm)	0.35	1.51	1.69	2.10
Al saturation	22.64	22.12	21.78	21.60
Fraction – sand (%)	55.23	55.21	55.21	55.10
Dust (%)	24.10	24.12	24.13	24.32
Clay (%)	20.67	20.67	20.66	20.58
Tekstur	Sandy clay loam	Sandy clay loam	Sandy clay loam	Sandy clay loam

rows of 25 cm. One week later, 50 g/pot. Then pots were given with basic fertilizers of Urea, TSP and KCl each dose of 0.25; 50 and 0.25 g/pot (50, 100 and 50 kg/ha). 7.5 mL *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* was mixed with 1 kg of soybean seeds. The mixture was then moistened with water, drained, mixed well, and stored in a shady place. It was left for 15 minutes and then was planted. The treatment of  $NH_4$ -molybdate as a source of Mo (containing 82% acid  $MoO_3$ ) (according to the treatment) was given at the planting time by dissolving each in 10 ml of water. The solution was sprinkled into polybags around the plants and then sprayed on the plants once every two weeks. After one week, the plants were thinned so only one plant/polybag was maintained until the plants were harvested. After the plants were harvested, the soil residue from the polybags was taken compositely at a depth of 0–25 cm to analyze its physical and chemical properties in the laboratory.

### Maintenance and harvest

Watering was performed daily by maintaining the groundwater condition at around 80% of field capacity. Thinning was carried out after the plants were 7 days old by removing one plant per polybag. Plant maintenance included embroidery, weeding, and hoarding. Plants were harvested after about three months of age. The characteristics of soybean plants that were ready to harvest were that, after all, the plant leaves were old or yellow.

### Observation and measurement

Vegetative growth observation included plant height, age of flower appearance, number of root nodules, percentage of effective root nodules, and nitrogen (N) and potassium (K) uptake. Meanwhile, observations for the generative phase were the weight of 100 seeds, the number of pods per plant, and dry seed weight per plant.

### Data analysis

The data from the experiment were statistically analyzed using SPSS version 16 software. The differences among treatments were compared using analysis of variance (ANOVA) with the least significant difference (LSD) posthoc test at a 5% probability level.

## RESULTS AND DISCUSSION

### Soil chemical and physical properties

The soil analysis results before (without treatment) and after the experiment (soil treatment results of mixed  $R_1$ ,  $R_2$  and  $R_3$ ) showed a change in the chemical properties of the soil. In general, there was a change in the chemical properties of the soil, i.e., pH, organic matter, total nitrogen (N), available phosphorus (P), potassium (K), Cation-exchange capacity (CEC), Ca, Mg, Mo, and Al. This was due to the treatment of *Rhizobium* strain

*Nod<sup>+</sup> Fix<sup>+</sup>*, supplementation of microelements Mo (NH<sub>4</sub>-molybdate), and lime CaCO<sub>3</sub> in ultisol soils which contributed to improving the chemical properties of the soil. Application of NH<sub>4</sub>-molybdate increased Mo from 0.35 ppm to 0.47 ppm. The application of lime CaCO<sub>3</sub> increased soil pH from 5.1 to 5.5. This occurred due to the ability of lime CaCO<sub>3</sub> to neutralize soil acidity through the mechanism of H<sup>+</sup> substitution by Ca<sup>2+</sup>. This condition provides a way for the availability of phosphorus freely through mineralization by soil biota in the solution. The basic reason for liming is to increase soil pH. As a result of changes in pH, changes in soil chemistry and biology occur, such as increased availability of phosphorus, nitrogen, potassium, calcium, and magnesium.

### Vegetative growth

The ANOVA results showed that cultivars (V) and the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate (R) had a significant effect on plant height, age of flower appearance, and number of root nodules, and percentage of effective root nodules (Table 2). There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate. The *Baturan* cultivar (V1) produced higher plant height, age of flower emergence, number of root nodules, and percentage of effective nodules. This was significantly different from the *Willis* cultivar (V2). These differences were due to the different characteristics of *Baturan* and *Willis* cultivars that affect photosynthetic activity, which determines plant height growth, flower appearance age, number of root nodules, and percentage of effective root nodules. Pregitzer et al. (2013) explained that some factors that influence plant growth and production are the environment, species or types of plants related to the genotype.

Table 2 shows that plant height, age of flower appearance, number of root nodules, and percentage of effective root nodules were increased with increasing doses of the combinations (CaCO<sub>3</sub>+NH<sub>4</sub>-molybdate). The highest increase was obtained in the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*+ CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/ha of 90.16 cm, 35.66 days, 57.22 seeds, and 79.76 (%), respectively. However, the results for the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*+ CaCO<sub>3</sub> 2.0 tons/ha+NH<sub>4</sub>-molybdate 750 g/ha were decreased statistically significantly. The increase in plant height, age of flower appearance, number of root nodules, and percentage of effective root nodules was supported by the availability of Mo for *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* by Mo administration. According to Schwarz and Mendel (2006), Mo is not only essential for plants, but also essential for microorganisms. Furthermore, more than 40 Mo enzymes are found in nature, most of which are found in bacteria, but only four Mo enzymes are found in plants. The increase in the availability of Mo in the soil has stimulated the growth, development, and activity of *Rhizobium* bacteria. Therefore, it increased the ability of *Rhizobium* bacteria to infect root nodules, resulting in a higher percentage of effective root nodules formed. Effective root nodules contain *Rhizobium* bacteria that can fix air N<sub>2</sub>.

Based on the results of Duncan's test in Table 2, the observation of plant height showed that the *Willis* cultivar achieved a higher plant height and was significantly different compared to the *Baturan* cultivar, both at the age of 2, 4, and 6 (weeks after planting) WAP, as well as significant differences in each treatment (R<sub>0</sub>, R<sub>1</sub>, R<sub>2</sub>, and R<sub>3</sub>). This was because the effectiveness of each treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>*, Lime (CaCO<sub>3</sub>), Molybdenum (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O) and Cobalt (CoC<sub>12</sub>) also depends on the type of variety used.

**Table 2.** Average plant height, age of flower appearance, number of root nodules, and percentage of effective root nodules on soybean cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate

Treatment		Plant height (cm)	Age of flower appearance (days)	Number of root nodules (seeds)	Percentage of effective root nodules (%)
Cultivars	V1	83.96 <sup>a</sup>	32.78 <sup>a</sup>	40.22 <sup>a</sup>	62.21 <sup>a</sup>
	V2	75.62 <sup>b</sup>	38.96 <sup>b</sup>	34.64 <sup>b</sup>	58.28 <sup>b</sup>
Sub-plots	R0	66.04 <sup>a</sup>	35.38 <sup>a</sup>	09.80 <sup>a</sup>	27.23 <sup>a</sup>
	R1	82.59 <sup>b</sup>	36.05 <sup>a</sup>	26.14 <sup>b</sup>	67.76 <sup>b</sup>
	R2	90.16 <sup>b</sup>	35.66 <sup>b</sup>	57.22 <sup>a</sup>	79.76 <sup>a</sup>
	R3	80.37 <sup>b</sup>	36.10 <sup>a</sup>	48.47 <sup>b</sup>	73.74 <sup>a</sup>

Note: Numbers followed by the same letter in each column are not significantly different at Duncan's test level =0.05.

One factor affecting growth and production is the species or types of plants related to genotype (Pregitzer et al., 2013).

The results of Duncan's test in Table 2 on the observation of the number of root nodules showed that different treatments gave different results on the number of root nodules. Treatment R<sub>2</sub> showed the highest number of root nodules. This was due to the suitability of the needs for bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>*, Lime (CaCO<sub>3</sub>), Molybdenum (NH<sub>4</sub>6Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O), and Cobalt (CoC<sub>2</sub>). Therefore, metabolic processes were increased due to increased assimilate formation, the influence of microelements Molybdenum and Cobalt, which play an essential role in enzyme systems (Bhattacharya et al., 2016). However, the soil analysis results before the study showed that Mo levels were quite low (0.97 ppm. In contrast, the requirement for soybeans was generally around 0.51 - 1.00 ppm (Ibañez et al., 2020). If the concentration of Mo in the soil is low, it will interfere with the formation of root nodules and inhibit N fixation by *Rhizobium* (Egamberdieva et al., 2018). This can be observed from the ability of plants to form effective root nodules. Ismunadji and Mahmud (1993) stated that applying Mo to soybean plants would increase the number of root nodules.

The increasing in the number of root nodules was caused by an increase in the number of nitrogenation enzymes and nitrate reductase, thereby increasing N fixation. (Imran et al., 2019) stated that Mo is the main component of the nitrogenation enzyme and nitrate reductase, whose preparation mechanism occurs due to a change in valence. Meanwhile, the process of forming root nodules occurs through cooperation between plants and N-fixing bacteria. Nodules that are active in symbiotic N binding, when split, will turn red due to the presence of leghaemoglobin. If the center of the root nodule is not red, it means it is not effective. Furthermore, (Zheng et al., 2020) and (Koskey et al., 2018) explained that root nodules are distinguished into effective and ineffective root nodules, in which the differences between the two root nodules are in size or shape, color, and location on the root.

The results of Duncan's test in Table 2 showed that the R<sub>2</sub> treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* + MoCo (1.0 : 0.6) kg/ha) produced the highest number of red nodules compared to other treatments. The high results indicated that the R<sub>2</sub> treatment impacted the formation of red root nodules. Therefore, it can infect plant roots in the

form of root nodules which are effective in fixing nitrogen from the air. This is presumably due to the ability of inoculation to meet the optimal N requirement for plant growth forming an effective red nodule. In addition, root nodules that are effective at fixing N can be recognized by their reddish color on the inside when the nodules are split. The red color is due to the presence of the pigment leghaemoglobin.

The observation results of the distribution pattern of the root nodules in Table 2 showed the best treatment of R<sub>2</sub>. The nodule distribution pattern determines the effectiveness of the nodule. Nodules scattered on the root neck indicate nodules effective in fixing N. The distribution pattern of root nodules explains the effectiveness of root nodules in fixing nitrogen free from the air. Effective root nodules are located on the taproot, while ineffective root nodules are scattered on lateral roots or root branches and are pale in color. The activity of N<sub>2</sub> fixation by bacteria is essential for overall N balance because the continuous form of fixed N can be lost due to denitrification and permeation (Howie & Echandi, 1983).

#### Nitrogen, phosphorus and potassium uptake

The ANOVA results showed that cultivars (V) had a significant effect on nitrogen (N) and potassium (K) uptake but had no significant effect on Phosphorus (P) uptake. The combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub> and NH<sub>4</sub>-molybdate (R) significantly affected N, P, and K uptake (Table 3). There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate. The *Baluran* cultivar (V<sub>1</sub>) produced higher N and K uptake, significantly different from the *Willis* cultivar (V<sub>2</sub>). However, the P uptake of treatments was not significantly different. According to

Table 3 shows that N, P, and K uptake increased with increasing doses of the combinations (CaCO<sub>3</sub> + NH<sub>4</sub>-molybdate). The highest increase was obtained in the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/ha of 852.7 mg/plant, 52.4 mg/plant, and 442.9 mg/plant, respectively. However, the results for the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/ha were decreased statistically significantly. The increased uptake of N, P, and K was thought to be the result of the ability of soybean cultivars to form root nodules in

**Table 3.** Average nutrient uptake of N, P and K in soybean cultivars and combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate

Treatment		Nutrient uptake (mg/plant)		
		N	P	K
Cultivars	V1	710.6 <sup>a</sup>	43.1 <sup>a</sup>	370.0 <sup>a</sup>
	V2	641.1 <sup>a</sup>	40.5 <sup>a</sup>	364.7 <sup>a</sup>
Sub-plots	R0	385.6 <sup>a</sup>	33.1 <sup>a</sup>	244.8 <sup>a</sup>
	R1	737.3 <sup>a</sup>	41.2 <sup>a</sup>	377.3 <sup>a</sup>
	R2	852.7 <sup>a</sup>	52.4 <sup>a</sup>	442.9 <sup>a</sup>
	R3	727.8 <sup>a</sup>	40.6 <sup>a</sup>	405.4 <sup>a</sup>

Note: numbers followed by the same letter in each column are not significantly different at Duncan's test level = 0.05.

response to *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* inoculation. In addition, it was supposed that the number of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* in that treatment was more suitable than others. The root nodules were formed if there was a suitability between *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* and its host.

The results of Duncan's test in Table 3 showed that the highest nitrogen uptake was found in the R<sub>2</sub> treatment compared to other treatments (R<sub>0</sub>, R<sub>1</sub> and R<sub>3</sub>). The significant effect of the R<sub>2</sub> treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* + MoCo (1.0 : 0.6) kg/ha) was thought to result from the soybean's ability to form root nodules in response to the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* inoculation. In addition, it is suspected that the number of bacterial strains *Nod<sup>+</sup> Fix<sup>+</sup>* in this treatment is more suitable than the other treatments, and effective root nodules would form if there were a suitability between the bacteria and its host.

#### Leaf color scale

The observation results of the leaf color scale in Table 4 showed that the treatment of bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* with Mo and CaCO<sub>3</sub> lime supplements on soybeans using ultisol soils affected the leaf color scale. Table 4 shows that the *Willis* and *Baluran* varieties treated with the *Nod<sup>+</sup> Fix<sup>+</sup>* bacterial strain, Mo and CaCO<sub>3</sub> lime supplements had various leaf color scales.

Table 4 showed that the results on the leaf color scale was the best treatment of R<sub>1</sub> for the *Baluran* cultivar compared to other treatments. This showed that the fixed N element was used to meet the N required by plants so the availability of N for plants increases. An effective fixation, 50–75% of the total N requirement by plants can be fulfilled. In general, this study showed the pattern of increasing plant height, root dry weight, number of root nodules, number of red nodules,

**Table 4.** Leaf color scale

Varietas	Treatments	Leaf color scales *			
		1	2	3	4
V <sub>1</sub>	R <sub>0</sub>	x			
	R <sub>1</sub>		x		
	R <sub>2</sub>			x	
	R <sub>3</sub>			x	
V <sub>2</sub>	R <sub>0</sub>	x			
	R <sub>1</sub>				x
	R <sub>2</sub>			x	
	R <sub>3</sub>			x	

Note: \* Criteria based on color scale, diagnosis of nutrient status from leaf color. The strengthening of legumes in relation to cropping system research project. Japan International Cooperation Agency (JICA): 1 – yellowish green, 2 – light green, 3 – green, 4 – slightly dark green.

N uptake, root nodule distribution pattern, leaf color scale, weight of 100 seeds, and dry seed yield. It was found that the treatment of the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* with supplements of microelements (MoCo), lime (CaCO<sub>3</sub>), and Cobalt, simultaneously increased the availability of Mo in the soil so as to increase the uptake of Mo by plants. Jaiswal et al. (2021) stated that the element Mo functions as an activator of nitrogenation and reducing enzymes which activate *Rhizobium* performance in the N fixation process so that N binding increases. Nitrogen can stimulate cell elongation, vegetative growth, enlarge pods, slow down the initiation phase, and increase seeds yield and content (Ohshima et al., 2013). The results of Duncan's test in Table 4 showed that the highest weight of 100 seeds was obtained in the R<sub>2</sub> treatment (bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* + MoCo (1.0 : 0.6) kg/ha) compared to other treatments (R<sub>0</sub>, R<sub>1</sub> and R<sub>3</sub>). This revealed that the treatment is vital for the growth and production of soybeans.

**Table 5.** The average weight of 100 seeds, number of pods per plant, and dry seed weight per plant

Treatment		Weight 100 seeds (g)	Number of pods per plant (piece)	Dry seed weight per plant (g)
Cultivars	V1	13.75 <sup>a</sup>	28.66 <sup>a</sup>	16.53 <sup>a</sup>
	V2	11.34 <sup>a</sup>	26.51 <sup>a</sup>	15.17 <sup>a</sup>
Sub-plots	R0	10.37 <sup>a</sup>	19.18 <sup>a</sup>	10.99 <sup>a</sup>
	R1	14.04 <sup>b</sup>	26.94 <sup>b</sup>	17.97 <sup>ab</sup>
	R2	14.49 <sup>b</sup>	35.89 <sup>b</sup>	19.45 <sup>b</sup>
	R3	11.88 <sup>a</sup>	28.34 <sup>a</sup>	15.00 <sup>a</sup>

Note: numbers followed by the same letter in each column are not significantly different at Duncan's test level = 0.05.

This was due to the Cofactor Mo (MoCo) forming the active site of all eukaryotic molybdenum enzymes.

### Yield components

The ANOVA results showed that cultivars (V) had no significant effect; while the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate had a significant effect on the weight of 100 seeds, number of pods per plant, and dry seed weight per plant. There was no interaction between the cultivars and the combinations of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*, CaCO<sub>3</sub>, and NH<sub>4</sub>-molybdate (Table 5).

Table 5 shows that the weight of 100 seeds, number of pods per plant, and dry seed weight per plant were increased with increasing doses of the combinations (CaCO<sub>3</sub>+ NH<sub>4</sub>-molybdate), the highest increase was achieved in the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>*+ CaCO<sub>3</sub> 1.5 tons/ha + NH<sub>4</sub>-molybdate 500 g/ha of 14.49 g, 35.89 pieces, and 19.45 g, respectively. However, the results for the combination of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/ha decreased statistically significantly. Increased vegetative growth continues in the generative growth of plants. The excess photosynthate, as a result of the photosynthesis process, was utilized to form pods, then stored in the form of seeds. In addition, the increase in N produced by plants will increase the synthesis of amino acids and proteins and plant organic compounds. These organic compounds are deposited on the seeds, increasing the dry seed weight per plant.

At the combination dose of *Rhizobium* strain *Nod<sup>+</sup> Fix<sup>+</sup>* + CaCO<sub>3</sub> 2.0 tons/ha + NH<sub>4</sub>-molybdate 750 g/ha, there was a tendency to decrease growth and yield of soybeans, presumably due to the excess of Mo and CaCO<sub>3</sub>. Based on Table

5, the highest dry seed yield was obtained in treatment R<sub>2</sub> and was significantly different compared to other treatments (R<sub>0</sub> and R<sub>3</sub>). This was because the R<sub>2</sub> treatment will increase nitrogen fixation to meet nitrogen needs for plants so that the availability of nitrogen for plants increases. In the R<sub>1</sub> treatment, there was a tendency to decrease the growth and production of soybeans, presumably due to excess (toxicity) of Mo and Co. The elements Mo and Co are micronutrients that are required in relatively low amounts with a narrow optimum range (Maggini et al., 2018). Therefore, its slightly excessive amounts will interfere with plant physiological processes and can even be toxic to plants. According to (Glass et al., 2012), the addition of excessive Mo will not affect the fixation of N.

The results of soil analysis before and after the study showed a change/increase in the content of physical/chemical properties of the soil, including parameters of soil pH, organic matter; N-total; Phosphor; Potassium; CEC; Ca; Mg; Mo and Co. This was due to the administration of the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>* with Micro (MoCo) and Lime (CaCO<sub>3</sub>) supplementations. In the podzolic marginal soil, the chemical properties were generally low/moderate (Table 1), and in each of these treatments, the *Nod<sup>+</sup> Fix<sup>+</sup>* strain bacteria, Lime (CaCO<sub>3</sub>), Molybdenum (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O, and Cobalt (CoC<sub>12</sub>) contributed to improving the chemical properties of the podzolic soil.

### CONCLUSION

This study showed that treating the bacterial strain *Nod<sup>+</sup> Fix<sup>+</sup>*+ MoCo (1.0 : 0.6) kg/ ha gave the best results on soybean growth and production, both for *Willis* and *Baluran* varieties on Ultisol marginal soils. Increasing the Mo dose will not significantly increase plant growth and yield.

## Acknowledgments

This work was supported by providing facilities from the Bosowa University, Indonesia.

## REFERENCES

- Al-Tawaha, A.R.M., McNeil, D., Othman, Y.A., AlRawashdeh, I.M., Adnan, M., Zamin, M., Qaisi, A.M., Al-Tawaha, A., Jahan, N., Shah, M.A., Khalid, S. 2022a. Legume Production and Climate Change. In *Improvement of Plant Production in the Era of Climate Change*. CRC Press, 221–248.
- Al-Tawaha, A.R.M.S., Khanum, S., Benkeblia, N., Khalid, S., Al-Tawaha, A.R., Mondal, M., Odat, N., Dey, A., Alimad, N., Thangadurai, D., Sangeetha, J. 2022b. *Adapting Crops to Climate Change. Climate Change and Agriculture: Perspectives, Sustainability and Resilience*, 53–77. <https://doi.org/10.1002/9781119789789.ch3>
- Al-Tawaha, A.R., Al-Tawaha, A.R.M. 2017. Response of soybean plants to exogenous application of yeast extract: Growth and chemical composition. *American-Eurasian Journal of Sustainable Agriculture*, 11(2), 31–36.
- Banaszkiewicz, T. 2011. Nutritional value of soybean meal. *Soybean and Nutrition*, 1–21. <https://doi.org/10.5772/23306>
- Bhattacharya, P.T., Misra, S.R., Hussain, M. 2016. Nutritional aspects of essential trace elements in oral health and disease: an extensive review. *Scientifica*, 1–12. <https://doi.org/10.1155/2016/5464373>
- Cieřla, J., Kopycińska, M., Lukowska, M., Bieganski, A., Janczarek, M. 2016. Surface properties of wild-type *Rhizobium leguminosarum* bv. *Trifolii* strain 24.2 and its derivatives with different extracellular polysaccharide content. *PLoS ONE*, 11(10), 1–21. <https://doi.org/10.1371/journal.pone.0165080>
- Cruz-Suárez, L.E., Tapia-Salazar, M., Villarreal-Cavazos, D., Beltrán-Rocha, J., Nieto-López, M.G., Lemme, A., Rieque-Marie, D. 2009. Apparent dry matter, energy, protein and amino acid digestibility of four soybean ingredients in white shrimp *Litopenaeus vannamei* juveniles. *Aquaculture*, 292, 87–94. <https://doi.org/10.1016/j.aquaculture.2009.03.026>
- Egamberdieva, D., Jabborova, D., Wirth, S.J., Alam, P., Alyemni, M.N., Ahmad, P. 2018. Interactive effects of nutrients and *Bradyrhizobium japonicum* on the growth and root architecture of soybean (*Glycine max* L.). *Front. Microbiol.*, 9, 1–11. <https://doi.org/10.3389/fmicb.2018.01000>
- Fitriatin, B.N., Yuniarti, A., Turmuktini, T., Ruswandi, F.K. 2014. The effect of phosphate solubilizing microbe producing growth regulators on soil phosphate, growth and yield of maize and fertilizer efficiency on Ultisol. *Eurasian J. Soil Sci.*, 3(2), 101–107. <https://doi.org/10.18393/ejss.34313>
- Glass, J.B., Axler, R.P., Chandra, S., Goldman, C.R. 2012. Molybdenum limitation of microbial nitrogen assimilation in aquatic ecosystems and pure cultures. *Front. Microbiol.*, 3, 1–11. <https://doi.org/10.3389/fmicb.2012.00331>
- Howie, W.J., Echandi, E. 1983. Rhizobacteria: influence type on plant growth and yield of potato. *Soil Biol. Biochem.*, 15(2), 127–132.
- Ibañez, T.B., Santos, L.F. de M., Lapaz, A. de M., Ribeiro, I.V., Ribeiro, F.V., Reis, A.R. Dos, Moreira, A., Heinrichs, R. 2020. Sulfur modulates yield and storage proteins in soybean grains. *Sci. Agric.*, 78(1), 1–9. <https://doi.org/10.1590/1678-992x-2019-0020>
- Imran, M., Sun, X., Hussain, S., Ali, U., Rana, M.S., Rasul, F., Saleem, M.H., Moussa, M.G., Bhandana, P., Afzal, J., Elyamine, A.M., Hu, C.X. 2019. Molybdenum-induced effects on nitrogen metabolism enzymes and elemental profile of winter wheat (*Triticum aestivum* L.) under different nitrogen sources. *Int. J. Mol. Sci.*, 20(3009), 1–17. <https://doi.org/10.3390/ijms20123009>
- Jaiswal, S.K., Mohammed, M., Ibnu, F.Y.I., Dakora, F.D. 2021. Rhizobia as a source of plant growth-promoting molecules: potential applications and possible operational mechanisms. *Front. Sustain. Food Syst.*, 4, 1–14. <https://doi.org/10.3389/fsufs.2020.619676>
- Koskey, G., Mburu, S.W., Kimiti, J.M., Ombori, O., Maingi, J.M., Njeru, E.M. 2018. Genetic characterization and diversity of *Rhizobium* isolated from root nodules of mid-altitude climbing bean (*Phaseolus vulgaris* L.) varieties. *Front. Microbiol.*, 9, 1–12. <https://doi.org/10.3389/fmicb.2018.00968>
- Lande, M., Bawankule, K.V., Solanki, R.D., Aware, R.G. 2019. Effect of bradyrhizobium broth on growth of root, shoot and nodule of soybean. *Int. J. Curr. Microbiol. Appl. Sci.*, 8(7), 1588–1596. <https://doi.org/10.20546/ijemas.2019.807.189>
- Maggini, S., Pierre, A., Calder, P.C. 2018. Immune function and micronutrient requirements change over the life course. *Nutrients*, 10(1531), 1–27. <https://doi.org/10.3390/nu10101531>
- Mengel, D., Ruiz-diaz, D., Asebedo, R., Maxwell, T. 2012. Nitrogen fertilization of nitrogen-stressed soybeans. *Better Crops*, 96, 14–15.
- Mitsch, M.J., Cowie, A., Finan, T.M. 2007. Malic enzyme cofactor and domain requirements for symbiotic N<sub>2</sub> fixation by *Sinorhizobium meliloti*. *J. Bacteriol.*, 189, 160–168. <https://doi.org/10.1128/JB.01425-06>
- Mmbaga, G.W.M., Mtei, K., Ndakidemi, P.A. 2015. Yield and fiscal benefits of inoculation supplemented with phosphorus (P) and Potassium (K) in Climbing Beans Grown in Northern Tanzania. *Agri. Sci.*,

- 6, 783–797. <https://doi.org/10.4236/as.2015.68076>
21. Ningrum, I.H., Irianto, H., Riptanti, E.W. 2018. Analysis of soybean production and import trends and its import factors in Indonesia. IOP Conference Series: Earth and Environmental Sci., 142, 1–8. <https://doi.org/10.1088/1755-1315/142/1/012059>
22. Ohyama, T., Minagawa, R., Ishikawa, S., Yamamoto, M., Phi Hung, N., Van, Obtake, N., Sueyoshi, K., Sato, T., Nagumo, Y., Takahashi, Y. 2013. Soybean Seed Production and Nitrogen Nutrition. A Comprehensive Survey of International Soybean Research - Genetics, Physiology, Agronomy and Nitrogen Relationships, 115–157. <https://doi.org/10.5772/52287>
23. Pregitzer, C.C., Bailey, J.K., Schweitzer, J.A. 2013. Genetic by environment interactions affect plant-soil linkages. Ecol. Evol. 3, 2322–2333. <https://doi.org/10.1002/ece3.618>
24. Rahman, M.A., Lee, S.H., Ji, H.C., Kabir, A.H., Jones, C.S., Lee, K.W. 2018. Importance of mineral nutrition for mitigating aluminum toxicity in plants on acidic soils: Current status and opportunities. International Journal of Molecular Sciences, 79(3073), 1–18. <https://doi.org/10.3390/ijms19103073>
25. Reuhs, B.L., Relić, B., Forsberg, L.S., Marie, C., Ojanen-Reuhs, T., Stephens, S.B., Wong, C.H., Jabbouri, S., Broughton, W.J. 2005. Structural characterization of a flavonoid-inducible *Pseudomonas aeruginosa* A-band-like O antigen of *Rhizobium* sp. strain NGR234, required for the formation of nitrogen-fixing nodules. J. Bacteriol., 187, 6479–6487. <https://doi.org/10.1128/JB.187.18.6479-6487.2005>
26. Salih, S., Hamd, S.A.M., Dagash, Y.M.I. 2015. The effects of rhizobium, mycorrhizal inoculations and diammonium phosphate (DAP) on nodulation, growth, and yield of soybean. Univers J. Agri. Res., 3, 11–14. <https://doi.org/10.13189/ujar.2015.030103>
27. Shiri Janagard, M., Ehadi-Segherloo, A. 2015. Inoculated soybean response to starter nitrogen in conventional cropping system in Moghan. J. Agron., 15, 26–32. <https://doi.org/10.3923/ja.2016.26.32>
28. Stiens, M., Schneiker, S., Pühler, A., Schlüter, A. 2007. Sequence analysis of the 181-kb accessory plasmid pSmeSM11b, isolated from a dominant *Sinorhizobium meliloti* strain identified during a long-term field release experiment. FEMS Microbio. Lett., 271, 297–309. <https://doi.org/10.1111/j.1574-6968.2007.00731.x>
29. Syahputra, E., Fauzi, Razali. 2015. The characteristics of the chemical properties of ultisols sub groups in some areas of Northern Sumatra. Theor. Appl. Climatol., 4, 1796–1803. <http://dx.doi.org/10.1016/j.ecolecon.2013.05.006>
30. Saranraj, P., Sivasakthivelan, P., Al-Tawaha, A.R.M., Bright, R., Al-Tawaha, A.R., Thangadurai, D., Sangeetha, J., Rauf, A., Khalid, S., Al Sultan, W., Safari, Z.S. 2021. Macronutrient management for the cultivation of Soybean (*Glycine max* L.): A review. In IOP Conference Series: Earth and Environmental Science. IOP Publishing, 788(1), 012055.
31. Wang, Q., Liu, J., Zhu, H. 2018. Genetic and molecular mechanisms underlying symbiotic specificity in legume-rhizobium interactions. Front. Plant Sci., 9, 1–8. <https://doi.org/10.3389/fpls.2018.00313>
32. Zheng, Y., Liang, J., Zhao, D.L., Meng, C., Xu, Z.C., Xie, Z.H., Zhang, C.S. 2020. The root nodule microbiome of cultivated and wild halophytic legumes showed similar diversity but distinct community structure in yellow river delta saline soils. Microorganisms, 8, 1–12. <https://doi.org/10.3390/microorganisms8020207>