

Substitution of sweet potato flour and corn starch to the growth, survival rate, feed conversion ratio and body chemical composition of juvenile *Litopenaeus vannamei*

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Abstract. For decades, the primary source of carbohydrates for vannamei shrimp (*Litopenaeus vannamei*) relied on the use of the more expensive corn starch. The combination of corn starch with sweet potato flour is expected to reduce the feed cost to a more affordable cost for the fish farmers. This study aimed to identify the most optimum substitution level between corn starch and sweet potato in the vannamei shrimp feed formulation. This study employed a completely randomized design with five formulations and three replications. The observed treatments were treatment A: 100% sweet potato flour and 0% corn starch, treatment B: 75% sweet potato flour and 25% corn starch, treatment C: 50% sweet potato flour and 50% corn starch, treatment D: 25% sweet potato and 75% corn starch, treatment E: 0% sweet potato flour and 100% corn starch. The results indicated that the combination of 50% sweet potato flour with 50% corn starch was the optimum treatment and the best body chemical composition, feed conversion ratio (FCR) and specific growth rate (SGR) were obtained. This study provides a scientific recommendation to the feed industry of vannamei shrimp and therefore, and the aim is to make the cost for the feed less expensive and affordable to the farmers.

Key Words: Aquaculture, chemical composition, feed, growth, substitution.

Introduction. Indonesia is a maritime country with a complex hydrography. Whiteleg shrimp (*Litopenaeus vannamei*) is one of the prominent fishery commodities with high export value (Ayisi et al 2017; Dastidar et al 2013; Haryotejo 2015; Hendradjat & Mangampa 2016; Statistics Indonesia 2016). Whiteleg shrimp is one type of crustacean farmed through aquaculture with higher productivity rate compared to other types of crustaceans including the giant tiger prawn (*Penaeus monodon*) (Liao & Chien 2011). South Sulawesi is one of the provinces in Indonesia that has great potential to be a whiteleg shrimp supplier. Although the export value of the whiteleg shrimp previously declined, in 2018, the total of whiteleg shrimp export value from South Sulawesi reached 4.462 tons, worth USD 42.8 million. There was an indication of decreasing shrimp production in South Sulawesi over the years. Therefore, in 2008, a national program entitled "Gerakan Kebangkitan Udang" (Shrimp Economic Revival Movement) was launched by the provincial government of South Sulawesi.

There were several factors contributing to the decrease of whiteleg shrimp production. One of the primary factors responsible for the failure in whiteleg shrimp farming in Indonesia was the inappropriate aquaculture technology and the inappropriate feeding according to the Water Environmental Carrying Capacity (WECC) (Zainuddin et al 2013; Zainuddin et al 2014). So far, feed cost contributed to 50% (Jatobá 2014), 58% (Son et al 2011) and 60-70% (Cummins et al 2013) of the production cost. Therefore, the government focused on the rearing, feeding, and nutrition development of whiteleg shrimp where feed cost could be potentially reduced (Lemos et al 2009; Tran et al 2013; Varadharajan & Pushparajan 2013).

Few studies have ventured into alternative feed development. The aim of alternative feed production is to minimize the use of animal protein substituted with plant protein (Davis & Arnold 2000; Divakaran et al 2000; Hernandez et al 2004; Tacon et al 2002). However, the use of both protein sources should be managed carefully. The excessive supplementation of protein would cause the expensive cost for feed production and produce waste that may degrade the water quality (Hari et al 2006; Lemos et al 2009; Wang et al 2015). Protein need for whiteleg shrimp's optimum growth, according to more sources, ranges from 30-44% (Cuzon et al 2004; Jatobá 2014; Wang et al 2015), 30-35% (Kumaran et al 2017) and above 30% (Kureshy & Davis 2002).

Several types of shrimp feed in South Sulawesi contained approximately 28-41% protein (Zainuddin et al 2014). Therefore, the use of excessive protein content in feed should be limited and optimized for growth (Bulbul et al 2015; Oujifard et al 2012). Other energy sources such as carbohydrates (protein-sparing effect by carbohydrates) with lower cost could be an alternative (Peres & Oliva-Teles 2002; Xia et al 2015; Zainuddin et al 2019). Many studies have reported the capable use of carbohydrates as an energy source (Cuzon et al 2004; Lee 2003; Zainuddin et al 2014). Carbohydrates have the potential to be an alternative low-cost energy source for shrimp growth (Niu et al 2012; Simon 2009; Wang et al 2014). So far, carbohydrate sources used in feed are microalgae or binders such as cassava flour, bran, wheat flour, or sago (Niu et al 2012; Wang et al 2015). Some studies have reported the use of corn starch in feed production (Guo et al 2006). However, less concern and emphasis were made to seek for the potential sources of carbohydrates used as shrimp feedstuff.

A study performed by Zainuddin et al (2017) indicated that in Indonesia, particularly in South Sulawesi, another available potential carbohydrate source is sweet potato flour. According to Zainuddin et al (2017) several carbohydrate sources such as sweet potato, bran, cassava, corn have a great potential to supply the carbohydrate component in shrimp feed since they contained lactose, glucose, and starch. Carbohydrates also contain oligosaccharide extracts that may serve as an aquaculture probiotic (Lesmanawati et al 2013; Theresia et al 2019). Therefore, in this study, the use of corn starch and sweet potato flour was performed to produce various less expensive carbohydrate sources. The use of alternative feed from local feedstuff as a carbohydrate source is expected to optimize potentially the growth of juvenile whiteleg shrimp. This study aimed to identify the optimum substitution level between corn starch and sweet potato in the vannamei shrimp feed formulation.

Material and Method

Experimental diets formulation. The production of formulated feed was initiated by collecting the feedstuff from local farmers in Maros, South Sulawesi. The employed formulated feedstuff contained 25% isoprotein. The sources of feedstuff consisted of fish meal, soy flour, shrimp cephalothorax flour, corn starch, sweet potato flour, vitamin, and mineral mix (Table 1). Formulated pelleted feed was dried to reduce its moisture content to 10%.

The proximate analysis encompassed crude protein content (CP), crude fat content (CF), water content, crude fiber content (CFib), nitrogen-free extract (NFE) and ash of the feed following the protocols suggested by the Association of Official Analytical Chemists (2005). The results of the feed proximate analysis are presented in Table 2 and the analysis was carried out in the Chemical Nutrition Laboratory, Faculty of Animal Science, Hasanuddin University.

Feed production was initiated by grinding all the collected feedstuff. All the collected ingredients were weighted according to the percentage and nutritional needs of *Litopenaeus vannamei*. All the ingredients were mixed and stirred to promote the homogeneity of the mixture. Fish oil, vitamin and mineral mix and warm water were mixed to produce a paste. The paste was converted to pellets by the feed pelleting machine.

Table 1
The composition and the percentage of experimental ingredients

Ingradiants	Treatment (%)				
Ingredients	Α	В	С	D	Ε
Sweet potato flour	0	10	20	30	40
Corn Starch	40	30	20	10	0
Fish meal	20	20	20	20	20
Cephalothorax flour	10	10	10	10	10
Soy flour	20	20	20	20	20
Fish oil	4	4	4	4	4
Vitamin mix	3	3	3	3	3
Mineral mix	3	3	3	3	3

Proximate composition of the feeds used in the treatments

Table 2

Treatment	Composition (%)						
	Water	СР	CF	CFib	NFE	Ash	
Α	6.37	25.99	11.88	2.44	49.40	10.41	
В	7.25	25.89	12.03	2.71	50.86	8.52	
С	6.58	25.38	11.98	2.39	52.41	7.81	
D	7.13	25.16	11.63	2.83	54.66	5.72	
E	7.32	25.46	11.38	2.91	55.54	4.70	

Feed trial and experimental design. The experiment was conducted on juvenile whiteleg shrimp with an initial individual weight of 0.77±0.01q. The shrimps were collected from the local shrimp nursery in Pangkep. The whiteleg shrimps were accommodated to the experimental formulated feed. Stocking density of the experimental animals used was 15 shrimps/tank. The tanks used in this study were glass tanks with a size of 60cmx50cmx50 cm and 150 liters volume each. The research took place from April to October 2017 in the mini hatchery of the Faculty of Marine and Fisheries Science, Hasanuddin University. The shrimp were acclimated in the hatchery for 1 week and reared for 8 weeks. We used 15 tanks that were filled with 120 liters of water each. We used sea water diluted with freshwater to obtain a salinity of 20 ppt. The percentage of feed provided to the whitelea shrimp was 10% of the juvenile body weight per day (Zainuddin et al 2014). The feeding was performed with a frequency of 4 times a day at the following hours: 06:00, 10:00, 14:00, 18:00 (Lestari et al 2019; Zainuddin et al 2014). The experimental formulated feed consisted of treatment A, B, C, D, and E with the following specifications: treatment A consisted of 100% sweet potato flour and 0% corn starch, treatment B consisted of 75% sweet potato flour and 25% corn starch, treatment C consisted of 50% sweet potato flour and 50% corn starch, treatment D consisted of 25% sweet potato and 75% corn starch, treatment E consisted of 0% sweet potato flour and 100% corn starch. Each treatment had 3 replications and was applied randomly to shrimp rearing media.

Statistical analysis. The observed parameters in this study encompassed the specific growth rate (SGR), survival rate (SR), feed conversion ratio (FCR) and body chemical composition of shrimp. By the end of the study, the bodyweight of the shrimps in each tank was calculated to measure their specific growth rate (SGR) and survival rate (SR) based on the method performed by Braga et al (2016), Niu et al (2012) and Zhang et al (2012). The amount of consumed feed during the rearing was calculated to identify its feed conversion ratio (FCR) based on the method performed by Hulefeld et al (2018). The formula for each calculation of the parameters is presented as follows:

Specific growth rate: SGR = 100 × (LnWt - LnW₀) / t

SGR = specific growth rate (%)
Wt = average weight of shrimp at time t (g)
Wo = average weight of shrimp in the initial research (g)
t = total days of culture period of research (day)

Survival rate: SR = Nt / No x 100

SR = survival rate (%) Nt = total survived shrimp No = total shrimp stocked

Feed conversion ratio (FCR): FCR = TFB / TB

TFB = total amount of feed broadcasted (g)
TB = total biomass of shrimp (g)

The body chemical composition was analyzed by proximate analysis at the end of the experiment by employing the AOAC method (AOAC 2005; Braga et al 2016; Dararat et al 2012). The chemical composition of the body shows crude protein (CP), crude fat (CF), crude fiber (CFib), nitrogen-free extract (NFE), ash, and glycogen (Glc).

Research data was statistically analyzed using the analysis of variance (ANOVA) using the SPSS software version 24. All results are expressed as average \pm standard deviation (SD). The influence of the different treatments was followed by the Tukey test at 95% confidence level. This test determines the difference of influence between treatments.

Results. Average weight gain of individual shrimp during the study is presented in figure 1. The pattern of all treatments had a linear tendency. It can be observed that the substitution of 50% sweet potato flour and 50% corn starch (treatment C) indicated the most significant treatment compared to all other treatments.

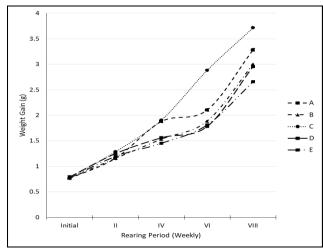


Figure 1. Body weight gain of juvenile whiteleg shrimp during the experiment.

The average rate of specific growth (SGR), survival rate (SR) and feed conversion ratio (FCR). The results of the analysis of variance showed that the treatment of substitution level of sweet potato flour and corn starch in the whiteleg shrimp feed could significantly affect (p<0.05) the specific growth rate and survival rate of juvenile whiteleg shrimp (Table 3). Means in the same column with different superscript letters ($^{a, b, c}$) are significantly different (p<0.05).

Table 3 The average rate of specific growth rate (SGR), survival rate (SR) and feed conversion ratio (FCR) of juvenile whiteleg shrimp during the experiment

Treatment	SGR (%/day)	Survival rate (%)	FCR
Α	3.61±0.25 ^b	67.00±0,00 ^b	1.90±0,10 ^b
В	3.57±0.40 ^b	$73.00 \pm 11,55$ ab	1.87±0,15 ^b
С	5.91±0.54ª	84.00±3,84ª	$1.40\pm0,10^{a}$
D	2.87±0.26 ^{bc}	67.00±0,00 ^b	1.93±0,06bc
E	2.51±0.31 ^c	67.00±0,00 ^b	1.93±0,12 ^c

Based on table 3, the substitution of 50% sweet potato flour and 50% corn starch was significantly different (p<0.05) from other treatments, while the substitution of 100% sweet potato flour and 0% corn starch, 75% sweet potato flour and 25% corn starch, and 25% sweet potato and 75% corn starch were not significantly different (p>0.05) in relation to the specific growth rate of juvenile whiteleg shrimp. The substitution of 25% sweet potato flour and 75% corn starch and the substitution of 0% sweet potato flour and 100% corn starch were also not significantly different (p>0.05) in the specific growth rate of juvenile whiteleg shrimp. Furthermore, the substitution of 50% sweet potato flour and 50% corn starch significantly different (p<0.05) from the other treatments except with a substitution of 75% sweet potato flour and 25% corn starch where the treatment was not significantly different (p>0.05) in the survival rate of juvenile whiteleg shrimp. The substitution formulation of 100% sweet potato flour and 0% corn starch, 75% sweet potato flour and 25% corn starch, 0% sweet potato flour 0% and 100% corn starch were not significantly different (p>0.05) in the survival rate of juvenile whiteleg shrimp.

Table 3 showed that the substitution of sweet potato flour and corn starch significantly affected (p<0.05) the feed conversion ratio. Table 3 indicated that the substitution of 50% sweet potato flour and 50% corn starch was significantly different (p<0.05) from the other treatments. The substitution of sweet potato flour 100% and corn starch 0% and the substitution of 75% sweet potato flour and 25% corn starch were not significantly different (p>0.05) from the substitution of 25% sweet potato flour and 75% corn starch. The substitution of 25% sweet potato flour and 75% corn starch was not significantly different (p>0.05) from the substitution of 0% sweet potato flour and 100% corn starch.

Body chemical composition. The results of body chemical composition analysis by the end of the experiment are presented in Table 4. Means in the same column with different superscript letters (a, b, c) are significantly different (p<0.05) and the following abbreviations were used: CP for crude protein (%), CF for crude fat (%), CFib for crude fiber (%), NFE for nitrogen-free extract (%), Glc for glycogen (g/100g sample).

Table 4 The body chemical composition of the experimental shrimp in final dry weight

Treatment	CP	CF	CFib	NFE	Ash	Glc
Α	76.22±0.76a	0.42±0.05ab	1.36±0.31	10.60±1.53 ^b	11.41±1.14 ^a	16.59±0.39 ^b
В	76.23±0.37ª	0.42 ± 0.04^{a}	1.31±0.04	6.05 ± 0.12^{a}	15.99±0.48 ^b	17.55±0.08 ^{cd}
С	78.33±0.14 ^b	0.40 ± 0.00 ab	1.06±0.02	9.67 ± 0.82^{ab}	10.54±0.97a	18.21±0.12 ^d
D	77.76±0.67 ^b	0.49 ± 0.02^{b}	1.19±0.14	9.40 ± 0.90^{ab}	11.16±0.38a	15.15±0.17ª
E	71.82±0.20 ^c	0.74±0.04 ^c	1.12±0.07	12.74±2.96 ^b	13.58±2.72ab	17.38±0.37 ^c

Based on table 4, the substitution of sweet potato flour and corn starch significantly (p<0.05) affected the body chemical composition of the experimental shrimp encompassing the crude protein, crude fat, nitrogen-free extract, ash, and glycogen content. In contrast, the substitution of sweet potato and corn starch did not significantly affect (p>0.05) the crude fiber. The protein content of whiteleg shrimp was higher with the substitution of 50% sweet potato flour and 50% corn starch compared to other treatments. The fat and ash content of experimental shrimp were significantly lower (p<0.05) in the substitution of 50% sweet potato and 50% corn starch compared to the other treatments. The body glycogen content resulted from the substitution of 50% sweet potato flour and 50% corn starch was significantly higher (p<0.05) compared to other treatments.

Discussion. The substitution of 50% sweet potato flour and 50% corn starch contributed to the most significant effect (p<0.05) because these two components contributed positively to the specific growth rate and survival rate of whiteleg shrimp. Both types of carbohydrate sources supplied a similar contribution to the specific growth rate and survival rate of whiteleg shrimp. According to studies performed by Krogdahl et al (2005), Niu et al (2012) and Simon (2009), different protein source could affect the shrimp and fish growth. The specific growth rate or survival rate of juvenile whiteleg shrimp would be reduced if the sweet potato flour percentage is increased to 75% and the corn starch is increased to 75% or 100%. The results can be described based on carbohydrate roles. The researchers assumed that the glucose and fructose content of sweet potato flour were higher compared to other carbohydrate sources (Rosas et al 2001; Theresia et al 2019). This is like starch content in corn starch compared to others. Consequently, the combination of 50%:50% of both produced better results. In relation to that, a few studies also mentioned that the alternative feed treatment with carbohydrate source could promote a significant effect on the juvenile whiteleg shrimp's growth (Amaya et al 2007; Davis et al 2002; Hasan 2001; Rosas et al 2001).

One determining factor in improving carbohydrate digestibility of the feed is the frequency of feeding (Zainuddin et al 2014). The results Zainuddin et al (2014) indicated that the feeding frequency of four times per day had a positive effect to the juvenile whiteleg shrimp growth and survival rate. The increase in growth was also affected by the feed factor and consumption behavior (Oujifard et al 2012). Growth can only occur when the consumed feed is higher than the needs of body maintenance. This is in line with the studies performed by Cuzon et al (2004) and Hasan (2001), indicating that the growth rate and survival rate are affected by the feed availability, age, water quality and water salinity (Diaz et al 2001; Hurtado et al 2006; Jannathulla et al 2019; Saoud et al 2003). The salinity level categorized as appropriate for whiteleg shrimp (Zainuddin et al 2019) ranged from 5-50 ppt (Hurtado et al 2006), 25-26.7 ppt (Diaz et al 2001) and 30-35 ppt (Saoud et al 2003). Braga et al (2016), Ghufron et al (2018), Soemardjati and Suriawan (2006) added that in addition to the feed quality, quantity, and the frequency, the supplied feed also affected the shrimp growth. The supplied feed should be appropriate and accurate to prevent feed deficiency (Hari et al 2006). Zainuddin (2014) stated that frequent feeding could upgrade the shrimp's capability of utilizing carbohydrate.

The highest feed efficiency was found in the substitution of 50% sweet potato and corn starch substitution. The lowest FCR was observed in the treatment of 50% sweet potato and corn starch substitution. FCR showed the conversion value of feed utilized by a reared animal to produce the body weight gain. The lowest FCR was found in the treatment where sweet potato flour and corn starch ratio was 50%:50%. The lowest generated FCR was 1.40, indicating that to produce 1 kg of shrimp, 1.40 kg of feed would be required. In comparison with other studies, the obtained FCR in this study was lower compared to the results (De Carvalho et al 2016; Qiu et al 2017; Olmos et al 2011;

Sookying & Davis 2011; Suárez et al 2009). This indicated that the supplied feed substitution contributed to the low conversion and was categorized as effective to improve reared shrimp biomass (Araneda et al 2008; Oujifard et al 2012).

The results of this study confirmed the argument that juvenile whiteleg shrimp could utilize higher amounts of carbohydrates. Although all the experimental feed from the treatments contained protein below 30%, the shrimp were able to utilize the feed (Tahe & Suwoyo 2011). A similar study was also performed by Hari et al (2006) with a protein content of 25% and 40%. The results indicated that the protein content of 25% with high carbohydrate content significantly promoted *Penaeus monodon*'s higher growth compared to the treatment of experimental feed with the protein content of 40%. In a study performed by Zainuddin et al (2014), it was identified that the use of feed with 30-50% carbohydrate content provided an equal contribution of glycogen and body chemical composition in juvenile whiteleg shrimp.

The use of carbohydrates in feed is particularly important due to some reasons: (a) carbohydrates are a cheaper energy source compared to the protein. Carbohydrate use could reduce feed cost that in turn, would decrease the total production cost (Simon 2009; Suárez et al 2009; Yang et al 2009). (b) At a certain level, carbohydrates could replace the energy from protein (*protein-sparing effect*) and due to its efficiency, the utilization of protein feed for growth can be improved (Wang et al 2015). (c) As a binder, carbohydrates (especially from certain feedstuff) could improve the physical quality of feed and reduce the percentage of feed dust (Son et al 2011). (d) As a nitrogen-free extract component, the supplementation of carbohydrates in feed could reduce the nitrogen waste to minimize the negative effect of feed on the environment (Dararat et al 2012; Olmos et al 2011).

Some results indicated similarities with a study performed by Hari et al (2006), stating that the carbohydrate source played an important role in reducing the ammonia content of nitrogen and in reducing the use of protein as an energy source (Braga et al 2016; Cuzon et al 2004; Samocha 2004). The quality of the carbohydrate relies on its sources. Sweet potato flour contains a high concentration of oligosaccharide (Viet et al 2018), glucose and fructose. Each component accounts for 4.49% and 4.23% respectively and has better content than other feedstuff sources such as cassava, sago, wheat, and corn (Zainuddin et al 2017).

The findings from the previous studies showed that the carbohydrate component in feed played an important role in boosting the metabolism, growth, and survival rate of shrimp or fish. Hari et al (2006) stated that the increase of growth was also affected by the feed factors, especially the inorganic nitrogen and carbohydrate content. Growth can only occur when the consumed feed is higher than the needs of body maintenance.

The results of this study indicated that the protein content of the experimental shrimp was identified in the treatment of 50% sweet potato and 50% corn starch substitution. This indicated the response of the protein-sparring effect by the carbohydrates (Rosas et al 2001). The results also showed that the treatment of 50% sweet potato flour and 50% corn starch substitution can lower the fat content of shrimps. The results indicated that the treatment of 50% sweet potato flour and corn starch substitution could deposit the glycogen significantly better compared to other treatments. The increase of carbohydrates and fat ration of feed was not linear with the increase of glycogen content in shrimps. The carbohydrates and fat ratio accounted for 4:5 in the substitution of 50% sweet potato flour and 50% corn starch were proven to deposit more glycogen compared to the carbohydrate and fat ratio accounted for 5:1 in the substitution of 0% sweet potato flour and 100% corn starch. The result of the study is different from the study performed by Li et al (2013) and Dong et al (2018), reporting that the glycogen content of juvenile blunt snout bream (Megalobrama amblycephala) increased as the ratio of the feeds carbohydrates and fat increased. The result of this study was like the result obtained by Ali & Jauncey (2004) stating that the glycogen content of African catfish was not significantly affected (p>0.05) by the increase of feed carbohydrate and lipid. The carbohydrate content in sweet potato flour and corn starch also affected the chemical glycogen content in shrimp's body. In the muscles, glycogen is a primary energy storage component that can amount to almost 2% of the shrimp total mass (Nur 2011).

Conclusion. Based on the results of the study, it can be concluded that the substitution of 50% sweet potato flour and 50% corn starch was categorized as the most effective treatment regarding the specific growth rate, survival rate, feed conversion ratio and chemical body composition of juvenile whiteleg shrimp. Further studies on the combinations of carbohydrates and proteins in whiteleg shrimp rearing is recommendable.

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References

- Ali M. Z., Jauncey K., 2004 Optimal dietary carbohydrate to lipid ratio in African catfish *Clarias gariepinus* (Burchell 1822). Aquaculture International 12(2):169–180.
- Amaya E., Davis D. A., Rouse D. B., 2007 Alternative diets for the Pacific white shrimp *Litopenaeus vannamei*. Aquaculture 262(2–4):419–425.
- Araneda M., Pérez E. P., Gasca-Leyva E., 2008 White shrimp *Penaeus vannamei* culture in freshwater at three densities: Condition state based on length and weight. Aquaculture 283(1–4):13–18.
- Association of Official Analytical Chemists (AOAC), 2005 Official Methods of Analysis (18th eds), Washington DC: Association of Official Analysis Chemists International.
- Ayisi C. L., Hua X., Apraku A., Afriyie G., Kyei B. A., 2017 Recent studies toward the development of practical diets for shrimp and their nutritional requirements. HAYATI Journal of Biosciences 24(3):109–117.
- Braga A., Magalhães V., Hanson T., Morris T. C., Samocha T. M., 2016 The effects of feeding commercial feed formulated for semi-intensive systems on *Litopenaeus vannamei* production and its profitability in a hyper-intensive biofloc-dominated system. Aquaculture Reports 3:172–177.
- Bulbul M., Kader M. A., Ambak M. A., Hossain M. S., Ishikawa M., Koshio S., 2015 Effects of crystalline amino acids, phytase and fish soluble supplements in improving nutritive values of high plant protein-based diets for kuruma shrimp, *Marsupenaeus japonicus*. Aquaculture 438:98–104.
- Cummins V. C., Webster C. D., Thompson K. R., Velasquez A., 2013 Replacement of fish meal with soybean meal, alone or in combination with distiller's dried grains with solubles in practical diets for pacific white shrimp, *Litopenaeus vannamei*, grown in a clear-water culture system. Journal of the World Aquaculture Society 44(6):775–785.
- Cuzon G., Lawrence A., Gaxiola G., Rosas C., Guillaume J., 2004 Nutrition of *Litopenaeus vannamei* reared in tanks or in ponds. Aquaculture 235(1–4):513–551.
- Dararat W., Lomthaisong K., Sanoamuang L., 2012 Biochemical composition of three species of fairy shrimp (*Branchiopoda: Anostraca*) from Thailand. Journal of Crustacean Biology 32(1):81–87.
- Dastidar P. G., Mallik A., Mandal N., 2013 Contribution of shrimp disease research to the development of the shrimp aquaculture industry: an analysis of the research and innovation structure across the countries. Scientometrics 97(3):659–674.

- Davis A., Arnold C. R., 2000 Replacement of fish meal in practical diets for the Pacific white shrimp, *Litopenaeus vannamei*. Aquaculture, 185(3–4):291–298.
- Davis D. A., Arnold C. R., McCallum I., 2002 Nutritional value of feed peas (*Pisum sativum*) in practical diet formulations for *Litopenaeus vannamei*. Aquaculture Nutrition 8(2):87–94.
- De Carvalho R. A. P. L. F., Ota R. H., Kadry V. O., Tacon A. G. J., Lemos D., 2016 Apparent digestibility of protein, energy and amino acids of six protein sources included at three levels in diets for juvenile white shrimp *Litopenaeus vannamei* reared in high performance conditions. Aquaculture 465:223–234.
- Diaz F., Farfan C., Sierra E., Re A. D., 2001 Effects of temperature and salinity fluctuation on the ammonium excretion and osmoregulation of juveniles of *Penaeus vannamei*, Boone. Marine and Freshwater Behaviour and Physiology 34(2):93–104.
- Divakaran S., Velasco M., Beyer E., Forster I., Tacon A. G. J., 2000 Soybean meal apparent digestibility for *Litopenaeus vannamei*, including a critique of methodology. Nutricion Acuicola, pp 267-276.
- Dong L.-F., Tong T., Zhang Q., Wang Q.-C., Xu M.-Z., Yu H.-R., Wang J., 2018 Effects of dietary carbohydrate to lipid ratio on growth, feed utilization, body composition and digestive enzyme activities of golden pompano (*Trachinotus ovatus*). Aquaculture Nutrition 24(1):341–347.
- Ghufron M., Lamid M., Sari P. D. W., Suprapto H., 2018 [The grow-out technique of vannamei shrimp (Litopenaeus vannamei) in the assisted pond of PT Central Proteina Prima Tbk in Randutatah Village, Paiton District, Probolinggo, East Java]. Journal of Aquaculture and Fish Health 7(2):70–77 [in Indonesian].
- Guo R., Liu Y.-J., Tian L.-X., Huang J.-W., 2006 Effect of dietary cornstarch levels on growth performance, digestibility, and microscopic structure in the white shrimp, *Litopenaeus vannamei* reared in brackish water. Aquaculture Nutrition, 12(1):83–88.
- Hari B., Madhusoodana Kurup B., Varghese J. T., Schrama J. W., Verdegem M. C. J., 2006 The effect of carbohydrate addition on water quality and the nitrogen budget in extensive shrimp culture systems. Aquaculture 252(2-4):248-263.
- Haryotejo B., 2015 [Analysis of market diversification of Indonesian Shrimp export commodity]. Jurnal Sosial Ekonomi Kelautan dan Perikanan 8(1):85–91 [in Indonesian].
- Hasan M. R., 2001 Nutrition and Feeding for Sustainable Aquaculture Development in the Third Millennium. Technical Proceedings of the Conference on Aquaculture in the Third Millennium, Bangladesh Agricultural University, Bangladesh, pp 193–219.
- Hendradjat E. A., Mangampa M., 2016 [Growth and survival rate of vannamei shrimp with traditional pattern plus in different densities]. Jurnal Riset Akuakultur 2(2):149–156 [in Indonesian].
- Hernandez C., Sarmiento-Pardo J., Gonzalez-Rodriguez B., Parra de la I. A., 2004 Replacement of fish meal with co-extruded wet tuna viscera and corn meal in diets for white shrimp (*Litopenaeus vannamei* Boone) Aquaculture Research 35(12):1153–1157.
- Hulefeld R., Habte-Tsion H.-M., Lalgudi R. S., McGraw B., Cain R., Allen K., Thompson K. R., Tidwell J. H., Kumar V., 2018 Nutritional evaluation of an improved soybean meal as a fishmeal replacer in the diet of Pacific white shrimp, *Litopenaeus vannamei*. Aquaculture Research 49(4):1414-1422.
- Hurtado M. A., Racotta I. S., Arjona O., Hernández-Rodríguez M., Goytortúa E., Civera R., Palacios E., 2006 Effect of hypo- and hyper-saline conditions on osmolarity and fatty acid composition of juvenile shrimp *Litopenaeus vannamei* (Boone, 1931) fed low- and high-HUFA diets. Aquaculture Research 37(13):1316–1326.
- Jannathulla R., Chitra V., Vasanthakumar D., Nagavel A., Ambasankar K., Muralidhar M., Dayal J. S., 2019 Effect of dietary lipid/essential fatty acid level on Pacific whiteleg

- shrimp, *Litopenaeus vannamei* (Boone, 1931) reared at three different water salinities Emphasis on growth, hemolymph indices and body composition. Aquaculture 513, 734405.
- Jatobá A., da Silva B. C., da Silva J. S., Vieira F. do N., Mouriño J. L. P., Seiffert W. Q., Toledo T. M., 2014 Protein levels for *Litopenaeus vannamei* in semi-intensive and biofloc systems. Aquaculture 432:365–371.
- Krogdahl Å., Hemre G. I., Mommsen T. P., 2005 Carbohydrates in fish nutrition: Digestion and absorption in postlarval stages. Aquaculture Nutrition.
- Kumaran M., Anand P., Kumar J. A., Ravisankar T., Paul J., Vasagam K. P. K., Raja K. A., 2017 Is Pacific white shrimp (*Penaeus vannamei*) farming in India is technically efficient? A comprehensive study. Aquaculture 468:262–270.
- Kureshy N., Davis D. A., 2002 Protein requirement for maintenance and maximum weight gain for the Pacific white shrimp, *Litopenaeus vannamei*. Aquaculture 204(1–2): 125–143.
- Lee S.-M., Kim K.-D., Lall S. P., 2003 Utilization of glucose, maltose, dextrin, and cellulose by juvenile flounder (*Paralichthys olivaceus*). Aquaculture 221(1–4):427–438.
- Lemos D., Lawrence A. L., Siccardi A. J., 2009 Prediction of apparent protein digestibility of ingredients and diets by in vitro pH-stat degree of protein hydrolysis with species-specific enzymes for juvenile Pacific white shrimp *Litopenaeus vannamei*. Aquaculture 295(1–2):89–98.
- Lesmanawati W., Widanarni, Sukenda, Purbiantoro W., 2013 [The potential of sweet potato oligosaccharide extract as a prebiotic of aquaculture probiotic bacteria]. Jurnal Sains Terapan 3(1):21–25 [in Indonesian].
- Lestari, Aslamyah S., Zainuddin, 2019 [Body chemical composition and glycogen levels at various doses of sweet potato (*Ipomea batatas*) as a prebiotic from Lactobacillus sp. in vannamei shrimp (*Litopenaeus vannamei*)]. Prosiding Simposium Nasional Kelautan dan Perikanan VI, Universitas Hasanuddin, Makassar, pp. 269-278 pp [in Indonesian].
- Li X. F., Wang Y., Liu W. B., Jiang G. Z., Zhu J., 2013 Effects of dietary carbohydrate/lipid ratios on growth performance, body composition and glucose metabolism of fingerling blunt snout bream *Megalobrama amblycephala*. Aquaculture Nutrition 19:701-708.
- Liao I. C., Chien Y.-H., 2011 The pacific white shrimp, Litopenaeus vannamei, in Asia: The world's most widely cultured alien crustacean. B.S. Galil et al. (eds.), In the Wrong Place Alien Marine Crustaceans: Distribution, Biology and Impacts, Invading Nature Springer Series in Invasion Ecology 6, pp. 489–519.
- Niu J., Lin H.-Z., Jiang S.-G., Chen X., Wu K.-C., Tian L.-X., Liu Y.-J., 2012 Effect of seven carbohydrate sources on juvenile *Penaeus monodon* growth performance, nutrient utilization efficiency and hepatopancreas enzyme activities of 6-phosphogluconate dehydrogenase, hexokinase and amylase. Animal Feed Science and Technology, 174(1–2):86–95.
- Nur, 2011 [The Effect of Various Combinations of Dietary Carbohydrate and Chromium Levels on Glycogen Deposits in the Hepatopancreas and Muscles of Tiger Shrimp (*Penaeus monodon*)]. Thesis Universitas Hasanuddin Makassar. Indonesia [in Indonesian].
- Olmos J., Ochoa L., Paniagua-Michel J., Contreras R., 2011 Functional feed assessment on *Litopenaeus vannamei* using 100% fish meal replacement by soybean meal, high levels of complex carbohydrates and bacillus probiotic strains. Marine Drugs 9(6):1119–1132.
- Oujifard A., Seyfabadi J., Abedian Kenari A., Rezaei M., 2012 Growth and apparent digestibility of nutrients, fatty acids, and amino acids in Pacific white shrimp,

- Litopenaeus vannamei, fed diets with rice protein concentrate as total and partial replacement of fish meal. Aquaculture 342–343:56–61.
- Peres H., Oliva-Teles A., 2002 Utilization of raw and gelatinized starch by European sea bass (*Dicentrarchus labrax*) juveniles. Aquaculture 205(3–4):287–299.
- Qiu X., Tian H., Davis D. A., 2017 Evaluation of a high protein distiller's dried grains product as a protein source in practical diets for Pacific white shrimp *Litopenaeus vannamei*. Aquaculture 480:1–10.
- Rosas C., Cuzon G., Gaxiola G., Le Priol Y., Pascual C., Rossignyol J., Contreras F., Sanchez A., Wormhoudt A. V., 2001 Metabolism and growth of juveniles of *Litopenaeus vannamei:* Effect of salinity and dietary carbohydrate levels. Journal of Experimental Marine Biology and Ecology 259(1):1–22.
- Samocha T., 2004 Substitution of fish meal by co-extruded soybean poultry by-product meal in practical diets for the Pacific white shrimp, *Litopenaeus vannamei*. Aquaculture 231(1–4):197–203.
- Saoud I. P., Davis D. A., Rouse D. B., 2003 Suitability studies of inland well waters for *Litopenaeus vannamei* culture. Aquaculture 217(1–4):373–383.
- Simon C. J., 2009 Identification of digestible carbohydrate sources for inclusion in formulated diets for juvenile spiny lobsters, *Jasus edwardsii*. Aquaculture 290(3–4):275–282.
- Soemardjati W., Suriawan A., 2006 [Technical guidelines for the cultivation of vannamei shrimp (*Litopenaeus vannamei*) in earthen pond]. Departemen Kelautan dan Perikanan Balai Budidaya Air Payau Situbondo, Indonesia [in Indonesian].
- Son V. N., Phuong N. T., Hai T. N., Yakupitiyage A., 2011 Production and economic efficiencies of intensive black tiger prawn (*Penaeus monodon*) culture during different cropping seasons in the Mekong Delta, Vietnam. Aquaculture International 19(3):555–566.
- Sookying D., Davis D. A., 2011 Pond production of Pacific white shrimp (*Litopenaeus vannamei*) fed high levels of soybean meal in various combinations. Aquaculture 319(1–2):141–149.
- Statistics Indonesia, 2016 Statistics of marine and coastal resources. In BPS statistics of Indonesia.
- Suárez J. A., Gaxiola G., Mendoz R., Cadavid S., Garcia G., Alanis G., Cuzon G., 2009 Substitution of fish meal with plant protein sources and energy budget for white shrimp *Litopenaeus vannamei* (Boone, 1931). Aquaculture 289(1–2):118–123.
- Tacon A. G., Cody J. J., Conqoest L. D., Divakaran S., Forster I. P., Decamp O. E., 2002 Effect of culture system on the nutrition and growth performance of Pacific white shrimp *Litopenaeus vannamei* (Boone) fed different diets. Aquaculture Nutrition 8(2):121–137.
- Tahe S., Suwoyo H. S., 2011 [Growth and survival rate of vannamei shrimp (*Litopenaeus vannamei*) with different feed combinations in controlled containers]. Jurnal Riset Akuakultur 6(1):31–40 [in Indonesian].
- Theresia M., Aslamyah S., Sriwulan, 2019 The utilization of sweet potatoes as prebiotics on the performance of *Lactobacillus* sp. in the vannamei shrimp digestion (*Litopenaeus vannamei*). International Journal of Environment, Agriculture and Biotechnology, 4(4):1087–1091.
- Tran N., Bailey C., Wilson N., Phillips M., 2013 Governance of global value chains in response to food safety and certification standards: The case of shrimp from Vietnam. World Development 45:325–336.
- Varadharajan D., Pushparajan N., 2013 Food and feeding habits of aquaculture candidate a potential crustacean of pacific white shrimp *Litopenaeus vannamei*, South East Coast of India. Journal of Aquaculture Research Development 4(1):161.
- Viet L. Q., Phu T. M., Q T. N., Hai T. N., 2018 The possibility of sweet potato (*Ipomoea batatas*) addition as feed for white leg shrimp (*Litopenaeus vannamei*) under biofloc

- rearing condition. International Journal of Scientific and Research Publications 8:2250–3153.
- Wang Li E. C., Wang S. F., Qin J. G., Chen X. F., Lai Q. M., Chen L. Q., 2015 Protein-sparing effect of carbohydrate in the diet of white shrimp *Litopenaeus vannamei* at low salinity. Aquaculture Nutrition 21(6):904–912.
- Wang Y. M., Wu F. J., Du L., Li G. Y., Takahashi K., Xue Y., Xue C. H., 2014 Effects of polysaccharides from abalone (*Haliotis discus hannai Ino*) on HepG2 cell proliferation. International Journal of Biological Macromolecules 66:354–361.
- Xia B., Gao Q.-F., Wang J., Li P., Zhang L., Zhang Z., 2015 Effects of dietary carbohydrate level on growth, biochemical composition, and glucose metabolism of juvenile sea cucumber *Apostichopus japonicus* (Selenka). Aquaculture 448:63–70.
- Yang Q., Zhou X., Zhou Q., Tan B., Chi S., Dong X., 2009 Apparent digestibility of selected feed ingredients for white shrimp *Litopenaeus vannamei*, Boone. Aquaculture Research 41(1):78–86.
- Zainuddin, Aslamiyah S., Tandipayuk H., 2013 [Increased production of vannamei shrimp (*Penaeus vannamei*) in South Sulawesi through the application of cheap, efficient, and environmentally friendly feed]. Penelitian Prioritas Nasional Masterplan Percepatan dan Perluasan Pembangunan Ekonomi Indonesia 2011–2025, Universitas Hasanuddin, Makassar, Indonesia [in Indonesian].
- Zainuddin, Aslamyah S., Hadijah, 2019 Effect of carbohydrate types on water quality, proximate composition, and glycogen content of white shrimp juvenile *Litopenaeus vannamei* (Boone, 1931) IOP Conference Series: Earth and Environmental Science, 253, 012003.
- Zainuddin, Aslamyah S., Haryati, 2017 Regional distribution map of carbohydrate producer and the feed material quality of vannamei shrimp in South Sulawesi. Applied Mechanics and Materials 862:132–137.
- Zainuddin, Haryati, Aslamyah S., 2014 Effect of dietary carbohydrate levels and feeding frequencies on growth and carbohydrate digestibility by white shrimp *Litopenaeus vannamei* under laboratory conditions. Journal of Aquaculture Research Development, 5(6):274.
- Zhang J., Liu Y., Tian L., Yang H., Liang G., Xu D., 2012 Effects of dietary mannan oligosaccharide on growth performance, gut morphology, and stress tolerance of juvenile Pacific white shrimp, *Litopenaeus vannamei*. Fish Shellfish Immunology 33(4):1027–1032.

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